


REVIEW OPEN ACCESS

Exercise Delays Brain Ageing Through Muscle-Brain Crosstalk

Shirin Pourteymour¹  | Rakesh Kumar Majhi^{2,3} | Frode A. Norheim¹ | Christian A. Drevon^{1,4}

¹Department of Nutrition, Institute of Basic Medical Sciences, Faculty of Medicine, University of Oslo, Oslo, Norway | ²Tissue Restoration Lab, Department of Biological Sciences and Bioengineering, Mehta Family Center for Engineering in Medicine, Indian Institute of Technology Kanpur, Kanpur, India | ³Center of Excellence in Cancer, Gangwal School of Medical Science and Technology, Indian Institute of Technology Kanpur, Kanpur, India | ⁴Vitas Ltd, Oslo, Norway

Correspondence: Shirin Pourteymour (shirin.pourteymour@medisin.uio.no)

Received: 21 November 2024 | **Revised:** 25 February 2025 | **Accepted:** 5 March 2025

Funding: Shirin Pourteymour has received funding from the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement No 801133.

Keywords: brain | brain ageing | CNS myelination | exercise | healthy ageing | myokines and brain health

ABSTRACT

Ageing is often accompanied by cognitive decline and an increased risk of dementia. Exercise is a powerful tool for slowing brain ageing and enhancing cognitive function, as well as alleviating depression, improving sleep, and promoting overall well-being. The connection between exercise and healthy brain ageing is particularly intriguing, with exercise-induced pathways playing key roles. This review explores the link between exercise and brain health, focusing on how skeletal muscle influences the brain through muscle–brain crosstalk. We examine the interaction between the brain with well-known myokines, including brain-derived neurotrophic factor, macrophage colony-stimulating factor, vascular endothelial growth factor and cathepsin B. Neuroinflammation accumulates in the ageing brain and leads to cognitive decline, impaired motor skills and increased susceptibility to neurodegenerative diseases. Finally, we examine the evidence on the effects of exercise on neuronal myelination in the central nervous system, a crucial factor in maintaining brain health throughout the lifespan.

1 | Introduction

1.1 | Exercise and the Brain

Understanding the mechanisms underlying brain ageing can aid in the prevention or even the reversal of progressive cognitive decline, such as in dementia. Dementia encompasses a broad group of disorders characterised by a gradual decline in cognition, memory loss, language deficits, visuospatial impairment, reduced executive function and alterations in mood or behaviour [1, 2]. Approximately 47 million people globally are living with dementia-related diseases, a figure projected to triple by 2050 [3]. Alzheimer's disease (AD), a prevalent form of dementia, affects approximately 6% of the population over

65 and becomes more common with age [4]. About 30% of AD cases can be attributed to modifiable risk factors, including hypertension, obesity, diabetes, and physical inactivity [3]. Dementia is associated with changes in the brain vasculature, size, morphology and signalling pathways [5]. Age-related atrophy of the grey matter [6–9], along with hippocampal shrinkage, is commonly observed and correlates with progressive memory loss [10, 11]. These changes may further contribute to a marked decline in learning capacity [12]. At the cellular level, synaptic contacts weaken, plasticity decreases [13], and hippocampal neurogenesis decreases [14, 15]. Although some degree of memory loss is a common consequence of ageing, it is not an inevitable outcome. The incidence of dementia increases with age, and cardiovascular diseases (CVD) enhance

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2025 The Author(s). *Cell Proliferation* published by Beijing Institute for Stem Cell and Regenerative Medicine and John Wiley & Sons Ltd.

the risk of cognitive impairment [16–19]. Thus, the prevention and treatment of CVD reduce the risk of dementia markedly [20]. This connection is demonstrated in mice with cardiac-selective overexpression of adenylyl cyclase type 8 (TGAC8), which exhibit elevated heart rate and contractility, along with altered neuroautonomic surveillance. These TGAC8 mice demonstrated significantly enhanced locomotor activity, evidenced by a 43% increase in distance travelled, a 38% increase in average speed and a 45% reduction in freezing time. Moreover, in the hippocampus of these mice, key neurotransmitter receptors are upregulated, indicating higher mental activity. More specifically, the brain perceives the increased myocardial humoral and functional output as a ‘sustained exercise-like’ scenario, prompting a response that activates central nervous system (CNS) output controlling locomotion. This response highlights how the heart–brain axis can play a significant role, with cardiovascular health being important for brain ageing [21].

There has been a significant shift in human lifestyle to a sedentary life over the past few centuries, contributing to the rise of lifestyle-associated diseases [22]. Exercise is essential for maintaining both metabolic and mental health [23]. The numerous health benefits of exercise are widely acknowledged, including its positive effects on obesity, type 2 diabetes, cardiovascular diseases, osteoporosis, depression, dementia, sleep disturbances, non-alcoholic fatty liver disease and various cancers [24–28]. Obesity, particularly in conjunction with social stress, impacts hippocampal structure and function, leading to reduced cognitive capacity partly due to diminished local pools of BDNF [29]. Exercise also reduces metabolic risk factors such as insulin resistance, blood lipid levels and chronic inflammation [30–32]. In contrast, 5 days of bed rest exert negative effects on muscle mass, insulin sensitivity, blood lipids and blood pressure [33]. Studies have shown that replacing sitting with standing during working hours or engaging in short bouts of light or moderate intensity walking between prolonged sitting may improve health outcomes, including increased plasma HDL levels and improved postprandial glucose and insulin levels [34, 35]. Exercise significantly influences human health, beginning in foetal development and continuing throughout the lifespan. Parental physical activity, both before and during pregnancy, influences the health of the mother and offspring [36, 37]. Maternal exercise during pregnancy is associated with benefits such as improved pregnancy outcomes, including reduced risk of macrosomia [38, 39], improved newborn neurobehavioural function [40] and cardiac autonomic health [41]. Furthermore, children of physically active mothers are more likely to adopt active lifestyles, lowering their risk of obesity and metabolic syndrome from infancy to adulthood [42]. Exercise during lactation also improves breastmilk composition, offering protection against obesity and inflammation [43] while supporting offspring brain development.

Large observational studies that track participants over time show that healthy adults who engage in regular exercise are less likely to develop dementia compared to inactive individuals [44, 45]. Exercise plays a crucial role in shaping our brain size, structure, and improves cognitive abilities [46]. A positive correlation between aerobic capacity and brain size has been reported [47]. Staying physically active throughout ageing

promotes CNS function and reduces neuroinflammation as well as the risk of developing neurodegenerative diseases [48]. A systematic review indicates that resistance exercise induces significant functional alterations in the brain, especially in the frontal lobe, promoting enhanced executive functions [49]. Exercise is also associated with reduced white matter atrophy and smaller volumes of white matter lesions [49]. Additionally, exercise positively impacts grey matter volume and cognitive function in late adulthood [50–53]. Age-related decline in cortical regions appears particularly responsive to exercise [54, 55]. An atlas of exercise-induced brain activation in mice reveals 255 brain regions activated by acute exercise, many of which were previously unlinked to exercise. Among these, 140 regions respond to both wheel and treadmill running, whereas 32 are unique to wheel running and 83 to treadmill running. Notably, forced treadmill running activates regions associated with stress, fear and pain [56].

Reduced blood–brain barrier (BBB) integrity in the human hippocampus is associated with early brain ageing and may be a contributing factor to cognitive impairment [57]. This may lead to hippocampal atrophy, which is also observed in AD [58, 59]. The hippocampus is known for its high degree of plasticity and neurogenesis, which provide an opportunity to enhance memory by improving hippocampal function [60]. For example, aerobic exercise has been shown to increase neurogenesis not only in the hippocampus but also in the hypothalamus and the subventricles [61–67]. Located along the lateral walls of the brain ventricles, the subventricular zone exhibits neurogenesis by producing new neurons in the adult brain [68]. A study of 115 individuals aged 50–70 years revealed that women who engaged in high levels of exercise had significantly larger volumes of the dorsolateral prefrontal cortex and temporal lobe compared to controls. Similarly, men who participated in high levels of exercise exhibited larger volumes in the temporal lobe [69]. The dorsolateral prefrontal cortex, which is positively influenced by exercise, plays a crucial role in cognitive functions like attention switching, working memory, rule maintenance, and inhibition of inappropriate responses [70] (Figure 1).

In old subjects, regular exercise significantly reduced brain tissue loss as compared to sedentary adults [71], and physically active old individuals have higher cognitive ability than sedentary old individuals of the same age [72–75]. Physical activity includes any energy-expending movement, whereas exercise is specifically structured and intentional, aimed at improving fitness. In this review, we use ‘exercise’ to denote purposeful physical activity.

2 | Potential Mechanisms of Action

Skeletal muscle has been identified over the past decades as a hub for the production, secretion and release of myokines, which are defined as secretory proteins. Myokines may function as hormones with local effects (autocrine or paracrine) or affect distant cells and organs through endocrine effects [76]. Muscles may communicate with the brain via extracellular vesicles [77], myometabolites such as lactate [78], enzymes like cathepsin B and amylase [79], and indirectly via other organs like the liver releasing the ketone body beta-hydroxybutyrate [80] (Figure 2).

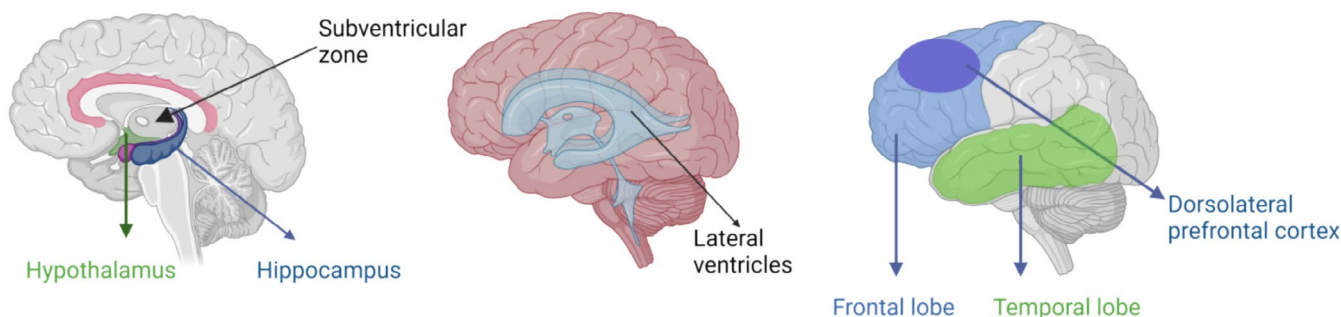


FIGURE 1 | Brain regions associated with exercise. Exercise promotes growth in areas of the subventricles, hypothalamus and dorsolateral prefrontal cortex. The hypothalamus plays a crucial role in managing emotions, regulating body temperature, and controlling basic needs like eating and sleeping. The hippocampus has a major role in storing and retrieving memories, ‘sending’ memories to appropriate sections of the cerebrum for storage and retrieval.

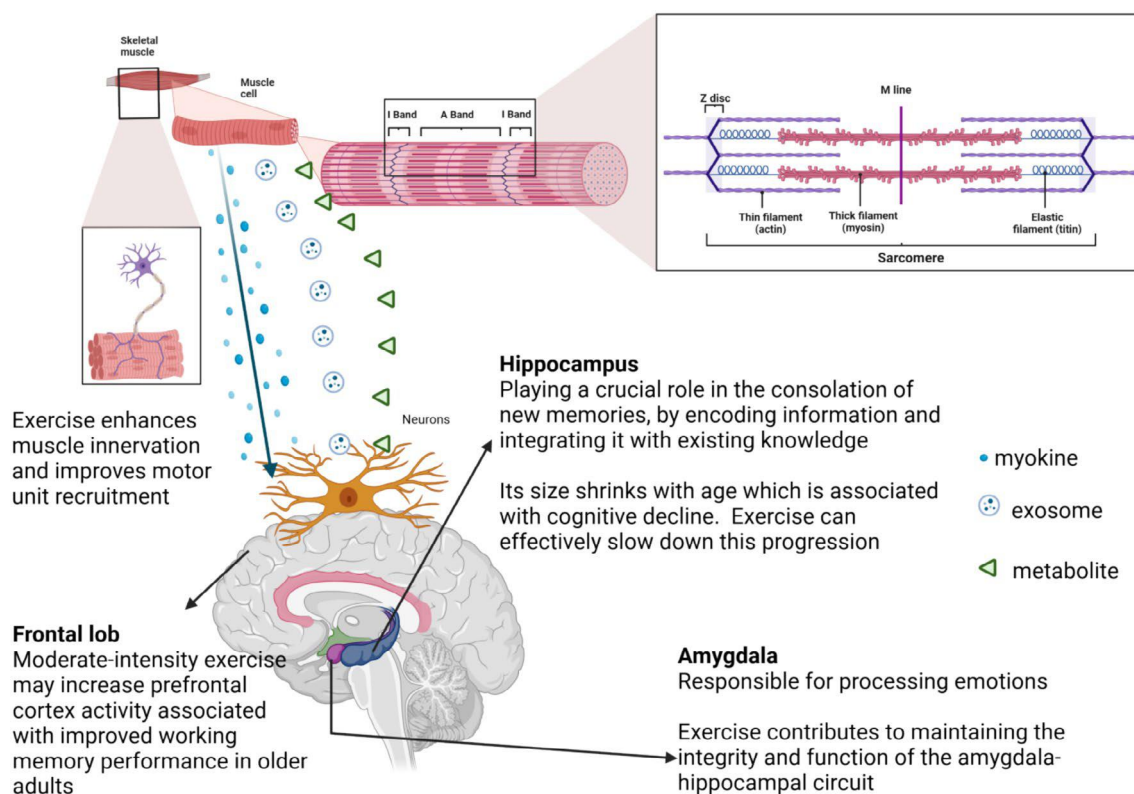


FIGURE 2 | Skeletal muscle communicates with other organs via nerves and secretory proteins (myokines), metabolites, and extracellular vesicles (exosomes) released into the extracellular space and the circulation [79]. Exercise helps preserve the integrity and function of the amygdala-hippocampal circuit [81]. Exercise-induced improvement in hippocampal function may promote better memory and cognition [73]. Moderate-intensity exercise may increase prefrontal cortex activity associated with improved working memory in older adults [82].

All these pathways can modulate cerebral variables such as blood flow, metabolic rate, mitochondrial biogenesis, neurogenesis, protein folding, oxidative effects, inflammation, and cell senescence, thereby affecting mood, sleep, cognition, food intake, neurodegeneration, and development of brain-related disorders [79].

We will examine selected myokines that may mediate the beneficial effects of exercise on brain health, particularly their roles in promoting neuronal myelination and regulating apolipoprotein E, a key factor in the accumulation of neuroinflammatory amyloid aggregates. Although irisin has been widely studied, we

will not include it in this review due to ongoing debate about its existence as an exercise-induced myokine with beneficial effects in humans [83, 84], as well as the availability of prior comprehensive reviews on this topic [85].

3 | Myokines

The fact that plasma transferred from exercised to sedentary animals improves cognitive functions supports the presence of exercise-induced circulatory factors [86, 87]. Exerkines are signalling molecules released in response to acute and/or chronic

exercise, acting via endocrine, paracrine or autocrine pathways. These factors are produced by a variety of organs, tissues, and cells, including skeletal muscle (myokines), heart (cardiokines), liver (hepatokines), white adipose tissue (adipokines), brown adipose tissue (baptokines), and neurons (neurokines) [76]. Several exercise-induced myokines have been described, including myostatin, interleukin-6 (IL6). Myostatin was probably the first myokine to be described [88], whereas IL6 is the most extensively studied myokine in response to exercise or muscle contraction [89, 90].

3.1 | Brain-Derived Neurotrophic Factor (BDNF)

BDNF (also called arbineurin) is an exerkine released by neurons, and partly also by skeletal muscle [91, 92] in response to exercise, especially aerobic activity. As a neurokine, BDNF highlights the systemic impact of exercise, benefiting both brain health and potentially other tissues through endocrine pathways [76]. It affects angiogenesis [93, 94], neuronal development, synaptic plasticity, growth and survival of neurons [95, 96]. BDNF is synthesised as a precursor protein (proBDNF, ~34 kDa) and later cleaved to the 18 amino acid signal peptides to generate proBDNF (14 kDa), which is then transported to the Golgi apparatus for conversion into mature BDNF (BDNF; ~14 kDa) by proteases [97, 98] (Figure 3A). In contrast to most growth factors, certain processing products of proBDNF and BDNF pro-peptide have biological functions often opposing those of the mature mBDNF [99, 100].

BDNF is one of the most highly expressed growth factors in the CNS [101]. It is expressed in several regions of the brain, particularly in the hypothalamus, cerebellum, amygdala, and the temporal lobe [102–105]. It binds to the tropomyosin-related kinase (TrkB) receptor, activates PI_3K , MAPK, PLC- γ , and GTPases of the Rho family, modulating synaptic plasticity [103], and enhancing dendritic growth and synaptic plasticity, and perhaps episodic memory [106–109], executive function, spatial memory, and learning [110–112].

Acute exercise significantly increases serum BDNF levels in concert with increasing exercise intensity [113–117]. Long-term exercise also increases basal BDNF levels in serum [118, 119]. Randomised clinical trials have shown that aerobic exercise in schizophrenic patients enhances neurocognition and BDNF levels by nearly 15% in comparison to patients on regular psychiatric treatment [120]. Serum BDNF levels increase in Parkinson's disease patients by 34% after 8 weeks of moderate interval training [121]. Moreover, serum BDNF concentrations were increased in obese individuals after 30 sessions of aerobic exercise [122].

Several studies have linked high levels of exercise-induced plasma BDNF to improved cognition [123–127]. Erickson et al. showed that a one-year aerobic exercise programme enhanced hippocampal volumes, memory, and serum BDNF levels [73]. A cross-sectional study assessed hippocampal volume, serum BDNF level and spatial memory in 142 participants (59–81 years of age), improved learning and memory in wild type and a Down syndrome mouse model [128]. Animal experiments indicated that BDNF may pass the BBB, and its blood levels may reflect the levels in the brain [129]. However, later studies have raised doubts about whether BDNF can pass the BBB or bind to its specific receptor on the BBB. Whether low, medium or high intensity exercise can affect the permeability of the BBB to BDNF remains to be ascertained. Notably, plasma concentration of mBDNF has been shown to increase during exercise [130, 131]. Mechanistic studies on its transport across the BBB are warranted to appreciate the beneficial effects of exercise-induced BDNF on brain structure and function.

Myocyte BDNF production plays a crucial role in mitigating the severe effects of myocardial ischemia [132]. In addition, the functional BDNF/TrkB signalling axis is essential for proper myocardial function, as underscored by recent studies revealing its importance for maintaining cardiac health under stressful conditions [104, 133]. Yang et al. explored whether myocardial BDNF/TrkB signalling influences cardiac responses to pathophysiological stress [134]. They reported a significant reduction in BDNF levels in heart failure (HF) mouse models as well as

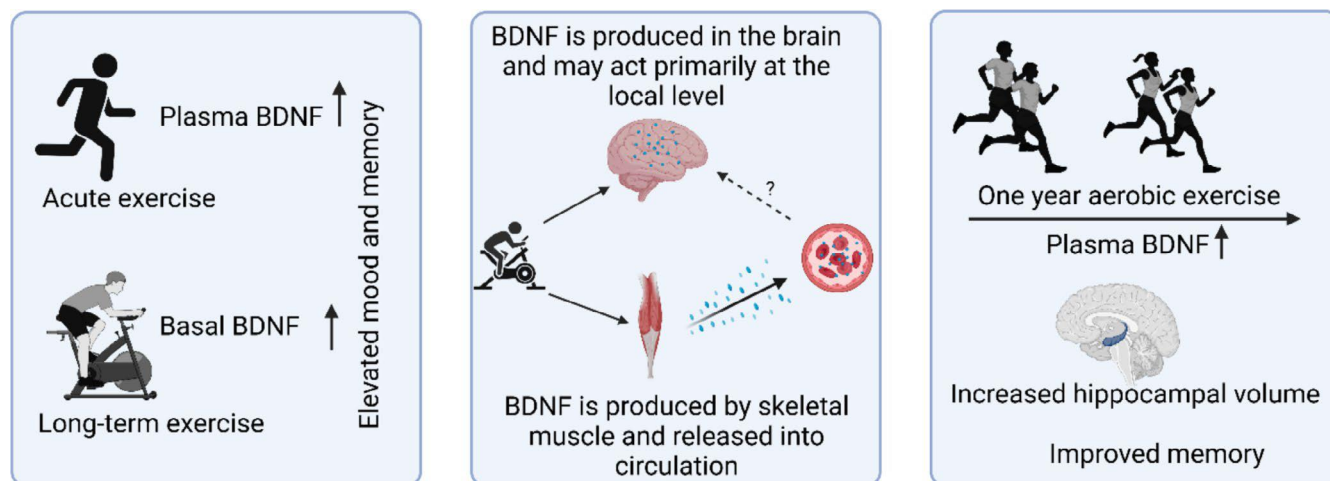


FIGURE 3 | The role of BDNF in mediating cognitive effects of PA. Acute and long-term exercise boost plasma BDNF levels, with long-term exercise further augmenting basal BDNF levels, suggesting a potential for sustained effects. Exercise enhances BDNF levels in the brain and blood, but the mechanism by which blood BDNF may influence the brain is unclear. A year-long exercise programme in older adults enhanced blood levels of BDNF, increased hippocampal volume, and improved memory performance [73].

humans with failing hearts. Interestingly, myocardial BDNF expression is increased in mice engaged in swimming exercise. Mice with a cardiac-specific TrkB knockout exhibited a compromised adaptive response to swimming. These findings highlight the critical role of myocardial BDNF in modulating cellular responses to swimming, positioning it as a potential therapeutic target for enhancing cardiac function in HF [134]. Furthermore, BDNF levels decline with age, which may contribute to the deterioration of cardiac function observed in elderly individuals, further emphasising the importance of BDNF in age-related cardiac pathophysiology [135].

3.2 | Macrophage Colony-Stimulating Factor 1

A series of transcripts regulated by acute and long-term exercise has been described recently, which identified macrophage colony-stimulating factor 1 (CSF1) as a secretory myokine up-regulated by acute exercise as well as long-term physical activity [136]. This was based on the measurement of CSF1 mRNA expression in skeletal muscle and its protein concentration in plasma before and after short- (hours) and long-term (12 weeks) exercise in 26 sedentary men [137]. We have also simulated skeletal muscle contraction by electrical-pulse stimulation *in vitro* and observed an increase in CSF1 mRNA in skeletal myocytes and CSF1 protein concentrations in the conditioned medium of cultured human skeletal myocytes [136]. These observations indicate that CSF1 is a secretory protein from skeletal muscle induced by exercise, and its production is increased in response to acute as well as long-term exercise (Figure 4) [139].

Studies in humans and mice show that CSF1 exerts different effects on the brain via microglia density and microglia distribution in both white and grey matter [138, 140]. Microglia are resident myeloid cells in the CNS acting both as glial cells, preserving homeostasis with trophic support to neurons and other glial cells, and as immune cells carrying functions important in response to tissue damage [141]. The CSF1 receptor (CSF1R) belongs to tyrosine kinase receptors and is activated by the two homodimer glycoprotein ligands: CSF1 [142] and IL-34 [143]. In contrast to CSF1, IL34 is not detectable in blood [144, 145], suggesting that its effects might be limited to its microenvironment. A specific CSF1R inhibitor given to mice led to a general

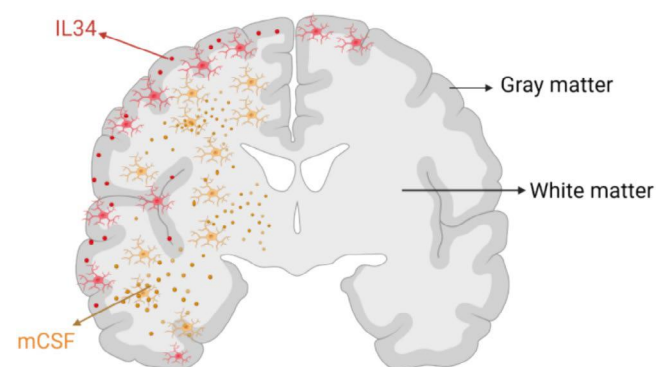


FIGURE 4 | Muscle-derived macrophage colony-stimulating factor (mCSF) is produced during exercise and remains elevated post-exercise. CSF1 affects a specific population of microglia in the brain in the white matter [138].

depletion of microglia in the brain, whereas blocking the CSF1 and IL34 function with antibodies specifically reduced microglia density in the white and grey matter, respectively [138]. These data suggest that CSF1 may be of importance for linking some effects of exercise to the brain.

3.3 | Vascular Endothelial Growth Factor (VEGF)

Skeletal muscle contributes to 60%–90% of peripheral vascular endothelial growth factor (VEGF) [146], and acute aerobic exercise transiently increases VEGF levels in muscle [147] and plasma [148, 149]. VEGF is a pro-angiogenic factor promoting vascularization in several tissue types and may play a role in neurogenesis. Intravenous administration of a VEGF antagonist attenuates running-induced hippocampal neurogenesis [150], whereas cerebroventricular administration of VEGF increases murine neurogenesis [151]. VEGF can cross the BBB, and most VEGF acting on the brain is probably derived from peripheral tissues [152]. By using a transgenic mouse deficient in skeletal muscle VEGF, it has been shown that VEGF deficiency reduces running-induced hippocampal neurogenesis [146].

3.4 | Interleukin 6 (IL6)

Contrary to CSF1, which maintains increased blood concentration also during long-term exercise (12 weeks and 4 times per week of combined endurance and strength training), muscle-secreted IL6 in plasma responds only to acute exercise and remains unchanged before and after 12 weeks of intervention [137]. Chronically elevated plasma IL6 in the resting state, on the other hand, seems to originate mostly from immune cells in adipose tissue [153]. The functions of IL6 include regulation of various biological processes associated with haematopoietic progenitor cells, adipose tissue, inflammation, hepatocytes, the placenta, the cardiovascular system, as well as the nervous and endocrine systems [154–156]. The expression of IL6 and IL6-R has been observed in both central and peripheral nervous tissues, including glial and neuronal cells and sympathetic and sensory ganglia [157–161]. Long-term excessive levels of IL6 may have negative effects on the homeostasis and chemistry of the nervous system. For instance, depressive patients and patients resistant to antidepressants have chronically elevated plasma IL6 concentrations in the CNS [162, 163]. Similarly, schizophrenic patients may have high plasma concentrations of IL6, and the severity of the disease is associated with the concentration of IL6 in the brain [164, 165]. Nevertheless, moderate and vigorous exercise may improve negative symptoms and brain function of schizophrenic patients [166].

3.5 | Cathepsin B

Cathepsin B is another exercise-responsive myokine [167] that may exert effects on the brain. Voluntary wheel running in cathepsin B knockout mice did not enhance hippocampal neurogenesis and spatial memory, opposite to the wild type, suggesting a role of cathepsin B in exercise-associated cognitive improvements [168]. Cathepsin B and BDNF seem to be linked to long-term exercise among men [169, 170], and cathepsin B may improve cognitive

function by increasing peripheral levels of BDNF [168, 171]. Cathepsin B may execute its effects on the brain via autophagy, modification of neuroinflammation, synaptic plasticity, neurogenesis and metabolic regulation [172, 173].

3.6 | Lactate

Although the brain only accounts for about 2% of the body weight, it receives ~20% of the total blood flow responsible for supplying oxygen, nutrients, hormones, and neurotransmitters, and removing carbon dioxide, toxins, and metabolic waste products [174]. Exercise-enhanced angiogenesis and density of microvessels in the brain of ageing rats are associated with improved brain function [175]. High-intensity exercise promotes anaerobic conditions in skeletal muscle, and glucose is shunted to glycolysis along with the release of lactate. Lactate can reach the brain and bind to a specific receptor named hydroxycarboxylic acid receptor 1 (HCAR1) on fibroblast-like cells lining pial blood vessels (intracranial blood vessels on the surface of the brain), and may induce the expression of VEGFa [78] known for its role in angiogenesis [176], neurogenesis, synaptic transmission, and plasticity [177]. Thus, lactate may serve as a mediator of skeletal muscle–brain communication in angiogenesis, enhancing cerebral blood supply to support optimal cognitive performance and brain function.

4 | Myelination

Exercise also may affect white matter in the brain; therefore, we will discuss how myelination is linked to cholesterol metabolism

and exercise (Figure 5). Myelination begins around birth in the peripheral nervous system, progressing to the spinal cord, and then in the brain throughout adulthood [178]. Myelination begins with the proliferation of oligodendrocyte precursor cells (OPCs) in the white matter. Then OPCs establish contact with axons and differentiate into myelinating oligodendrocytes.

Cholesterol increases myelin viscosity and stabilises myelin lipids and proteins, making it a critical and limiting factor for the development of myelin membranes in the CNS [179, 180]. BBB restricts the entry of peripheral cholesterol, meaning the primary source of brain cholesterol is local de novo synthesis in oligodendrocytes or astrocytes [181, 182]. Cholesterol in the CNS has a long half-life, lasting about 1 year in mice and up to 5 years in humans, compared to just a few days in plasma [183, 184].

4.1 | Exercise and Myelination in Rodents

Several studies have shown that exercise promotes myelination in rodents. Long-term running exercise has been found to stimulate myelination in the motor cortex [185, 186], reverse toxin-induced demyelination [187] and preserve myelinated fibres in brain white matter [188]. These findings are supported by a systematic meta-analysis of 21 articles demonstrating the positive impact of exercise on myelin sheath regeneration in rodents [189].

A group of ‘depressive’ rats exhibited reduced length and volume of myelinated fibres, along with reduced volume and thickness of myelin sheaths [190]. However, the quality of myelin

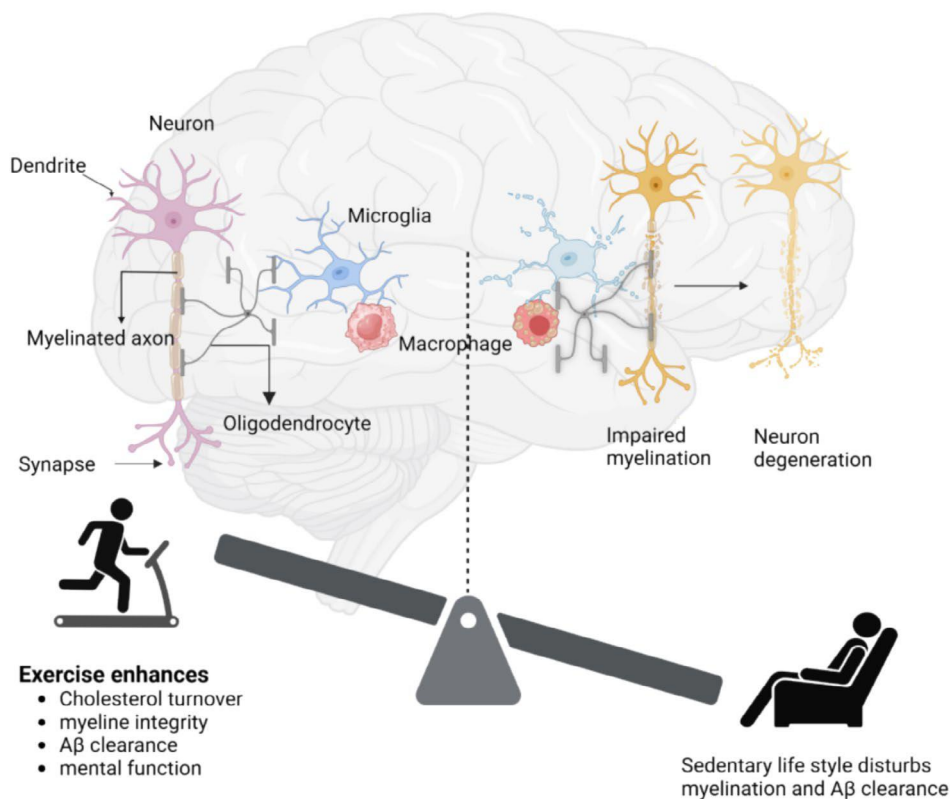


FIGURE 5 | Physical activity may be beneficial for myelin formation and cholesterol homeostasis in the CNS through oligodendrocyte cells, which play an important role in both the synthesis and recycling of cholesterol required for the function of cell membranes and myelination.

improved significantly following exercise compared to a control group [190]. Similarly, AD mice on a running programme for 4 months showed enhanced learning and spatial memory, as well as increased volumes of myelinated fibres in the CA1 region of the hippocampus [191].

4.2 | Exercise and Myelination in Humans

A study including 88 healthy, untrained adults aged 60–78 years showed that engaging in exercise and avoiding sedentary behaviour enhanced myelin thickness in the brain [192]. Another study involving 10 sedentary subjects (74.5 ± 4.3 years of age) and 10 elite athletes (aged 72.2 ± 5.3 years, endurance training > 15 years) reported that life-long exercise was associated with smaller lesions in brain white matter and better motor control and coordination [193]. Voss et al. examined the impact of a one-year aerobic fitness intervention on the integrity of cerebral white matter and cognitive function in 70 adults aged 55–80 years [194]. The fitness program did not improve white matter integrity or cognitive function on a group level, but individuals with a higher score of aerobic fitness in the program experienced improved white matter integrity in certain brain regions and enhanced short-term memory [194]. This study indicated that improved white matter integrity was not directly correlated with memory improvement but was associated with the extent of fitness gain [194]. Similar results were observed in a one-year randomised controlled trial of aerobic exercise involving 36 patients with amnesic mild cognitive impairment [195]. A systematic review of 38 studies comparing cognitive and exercise training showed that cognitive training improved white matter microstructure, whereas exercise tended to enhance connectivity and larger structural outcomes concerning grey and white matter [196]. Some studies have reported a lack of correlation between exercise and myelination [197, 198]. However, it is important to note a limitation in these studies, as participants self-reported their exercise levels through monthly telephone interviews [199].

4.3 | Apolipoprotein E, Myelination, and Exercise

Apolipoprotein E (ApoE) is important in cholesterol homeostasis, with high expression in the liver and brain [200–202], and ApoE is associated with AD [203, 204]. In aged ApoE-knockout mice, foamy glia cells (lipid-loaded) accumulate in regions like the *thalamus*, *fimbria hippocampi*, and *hippocampus* associated with neurodegenerative and behavioural changes [205]. Long-term aerobic exercise from midlife continued into old age mitigated neurovascular decline, reduced neuroinflammation, and enhanced synaptic plasticity and behavioural capabilities in mice [206]. The protective effects of exercise were linked to the preservation of astrocytic ApoE levels [206]. However, exercise had little effect on neurovascular decline or microglia activation in the absence of ApoE, suggesting that exercise may stabilise ApoE function [206].

ApoE2, ApoE3, and ApoE4 [207] represent common polymorphisms in the APOE gene; they play a significant role in lipid metabolism and CVD [208]. ApoE3 is the most prevalent variant in the general population, whereas ApoE4 constitutes the

most significant genetic risk factor for AD [209]. These single amino acid polymorphisms modify the structure and function of ApoE, influencing its binding to lipids as well as receptors [210]. Prevalence varies by region, with APOE4 more common in certain populations, like Central Africa (40%), and less so in others, like South China (less than 10%). There is a gradient of APOE4 distribution in Europe and Asia, with higher prevalence in northern Europe and Asia (ca. 25%). APOE2 prevalence is higher in Africa and Oceania, with 9.9% and 11.1% penetrance, respectively. This variation may suggest selective advantages for specific alleles in different climates and populations [211–213].

Blanchard et al. [214] examined 32 post-mortem brains (12 *ApoE3/3*, 12 *ApoE3/4* and 8 *ApoE4/4* carriers), 20 with AD, and the total group included 20 ApoE4 carriers. Using single-nucleus transcriptional profiling, they identified affected pathways related to cholesterol synthesis in oligodendrocytes (ODC) in ApoE4 carriers. Analysis of the hippocampus and prefrontal cortex revealed cholesteryl ester accumulation in *ApoE4* carriers and reduced myelination, suggesting issues with cholesterol incorporation into myelin [214]. They also investigated the impact of ApoE4 on oligodendrocytes by creating human oligodendrocytes from induced pluripotent stem cells with engineered ApoE4 or ApoE3. They observed high cholesterol accumulation in ApoE4-carrying oligodendrocytes, particularly around the endoplasmic reticulum (ER). This cholesterol accumulation induced ER stress and promoted nuclear translocation of the stress-activated transcription factor ATF6 [214]. Promoting cholesterol transport with cyclodextrin reduced cellular cholesterol accumulation, potentially incorporating cholesterol into myelin [214]. These results are compatible with dysfunctional myelination in asymptomatic *ApoE4* carriers and reduced myelin levels in infants with ApoE4 [215, 216].

Amyloid-beta ($A\beta$) is predominantly synthesised in neurons by proteolytic cleavage of amyloid precursor protein. The brain employs multiple pathways for removal of $A\beta$, including (a) cellular uptake and degradation; (b) enzymatic degradation; (c) clearance via the BBB; (d) clearance via interstitial fluid (ISF) bulk flow and (e) the glymphatic pathway. Cellular uptake of $A\beta$ is facilitated by receptors like LDL receptor-related protein 1 (LRP1), LDL receptor (LDLR), and heparan sulphate proteoglycan (HSPG) [217]. ApoE, primarily synthesised and lipidated by astrocytes, plays a crucial role in the clearance of $A\beta$. A subpool of ApoE-containing lipoprotein particles interacts with soluble $A\beta$ released from neurons into the brain ISF. Elimination of soluble $A\beta$ from brain ISF takes place in an ApoE isoform-dependent manner, where ApoE4 displays lower efficacy compared to ApoE2 or ApoE3 [218]. Abnormal myelination in the hippocampus may occur even before aggregation of amyloid and tau in AD mice [219]. It is possible that ApoE4 may compromise cholesterol efflux/metabolism, promoting impaired myelination, which may induce cholesterol accumulation in ODC, transforming them into foam cells.

In a murine model of multiple sclerosis, it was shown that exercise may improve BBB integrity and influence brain cholesterol homeostasis [220]. Scientists have reported effects of exercise on cholesterol flux in animals [221, 222]; thus, exercise may stimulate cholesterol efflux from brain cells, mitigating formation of foam cells. This may support optimal function of ODC, glia cells

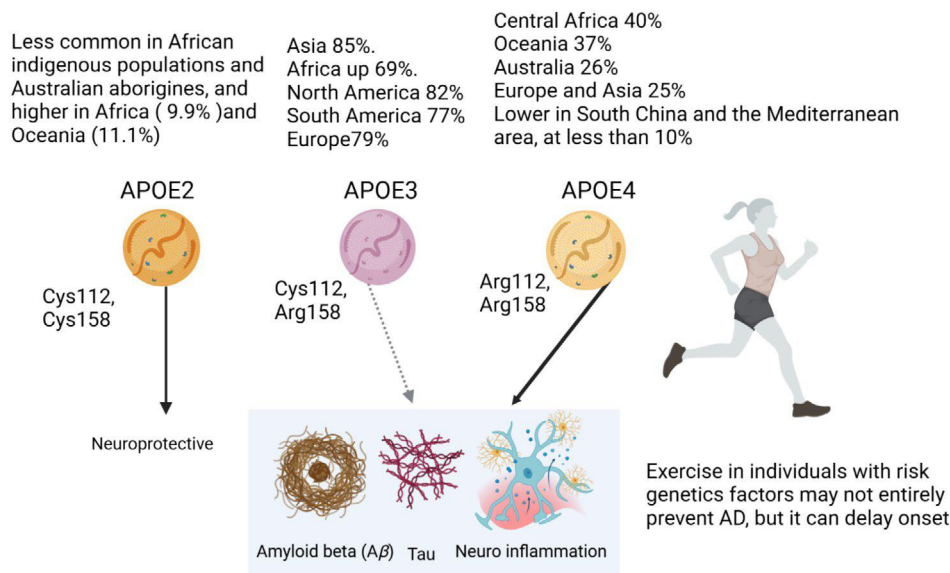


FIGURE 6 | APOE alleles and their effects on CNS. APOE3 is the most common isoform, making up about 80% of alleles globally. APOE2 and APOE4 are less common, constituting around 5%–10% and 10%–15%, respectively.

and phagocytes, enhancing cholesterol turnover, improving myelination, and clearing A β (Figure 6).

5 | Effect of Exercise on the Blood–Brain Barrier

The concept of BBB dates back to the late 19th century, founded on observations that dyes and biologically active substances did not impact the brain or behaviour unless directly injected into the CNS [223, 224]. The BBB is dynamic throughout life; for instance, the transport of amino acids by the BBB varies significantly from neonates to adults [225]. A recent study on 20,000 subjects explored the permeability of the blood–cerebrospinal fluid barrier and the BBB, revealing significant sex differences in barrier integrity across all age groups indicated by the (Cerebrospinal fluid) CSF/serum albumin ratio, a key biomarker of barrier function [226]. These findings indicated that males generally exhibit lower barrier integrity. Moreover, CSF reabsorption slows with age, contributing to higher CSF/blood albumin ratios. The extent to which this age-related increase in albumin ratios is due to BBB leakage or reduced CSF reabsorption remains unresolved [227].

Imaging, especially dynamic-contrast-enhanced magnetic resonance imaging (DCE-MRI), is a preferred method for assessing BBB function in humans. Even during healthy ageing without pathological cognitive decline, BBB disruption is evident in the hippocampus, as well as in grey and white matter [228]. This disruption correlates with cognitive decline often associated with healthy ageing, particularly in delayed recall [229], highlighting a connection between BBB integrity and certain cognitive changes associated with ageing.

Animal studies consistently show beneficial effects of physical activity on BBB structure and function [230, 231]. Heart failure is known to compromise BBB integrity, significantly contributing to autonomic nervous system dysfunction by increasing caveolin-1 expression, enhancing vesicle trafficking and weakening

tight junctions, thereby elevating BBB permeability [232]. This BBB dysfunction disrupts neural regulation and exacerbates the systemic effects of HF, whereas exercising Wistar rats restores BBB integrity by normalising caveolin-1 expression, reducing vesicle trafficking and strengthening tight junctions, all lowering BBB permeability [232].

Insulin resistance within the CNS is often associated with cognitive impairments like AD (ref). A study with CD-1 male mice showed that acute exercise increased insulin transport across the BBB and improved insulin vascular binding in the brain for both sexes [233].

A study demonstrates that long-term exercise enhances amyloid- β clearance by improving BBB function in 5XFAD mice, a transgenic model of AD that overexpresses amyloid- β (A β). Exosomes derived from exercised 5XFAD mice promote proliferation and upregulate the mRNA expression of PDGFR β , ZO-1, and claudin-5 in primary brain pericytes and endothelial cells in vitro. PDGFR β is essential for pericyte survival and BBB maintenance, while ZO-1 and claudin-5 are key tight junction proteins that regulate endothelial barrier integrity and selective permeability. Notably, these exosomes exhibit significant alterations in miR-532-5p levels, and when administered to sedentary mice or transfected into primary brain cells, they replicate the BBB improvements observed in exercised mice. These findings suggest that exercise-induced exosome signalling enhances BBB function by stabilising pericytes and reinforcing tight junction integrity, which may contribute to improved amyloid- β clearance and neuroprotection in AD [234].

The effects of aerobic exercise on BBB integrity were also evaluated in a rat model of multiple sclerosis, demonstrating that exercise may improve markers of BBB integrity and reduce neuronal apoptosis [235].

A human intervention study explored the anti-inflammatory effects of exercise and taurine supplementation on BBB integrity,

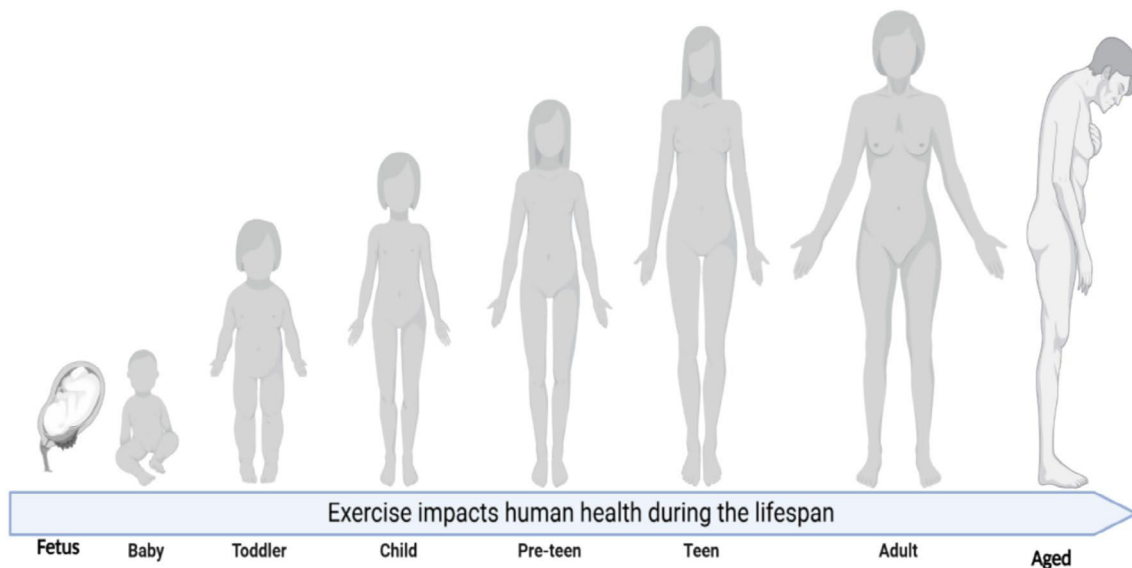


FIGURE 7 | Exercise influences human health from foetal development to advanced age, highlighting the importance of physical activity throughout life. The benefits begin with maternal exercise, which positively impacts foetal development and extends into offspring health. Children born to physically active mothers tend to experience better health outcomes and are more likely to age healthily compared to those born to sedentary mothers.

inflammation markers, and cognitive function in 48 elderly women for 14 weeks. Participants were divided into groups: (a) combined exercise training; (b) taurine supplementation; (c) both exercise and taurine or (d) a control group with no intervention. Exercise alone (a), as well as in combination with taurine (b), reduced inflammation and preserved BBB integrity. Importantly, the group receiving both interventions (c) exhibited a significant improvement in cognitive function by scores on the Mini-Mental State Examination [236].

A 12-week moderate-intensity aerobic exercise program in 56 methamphetamine-dependent individuals (aged 18–45) significantly improved neurofilament light chain and neuron-specific enolase blood levels compared to standard detoxification, indicating enhanced neurological recovery and blood–brain barrier integrity in the exercise group [237].

The reviewed articles consistently demonstrate that exercise improved the structure and function of the BBB, irrespective of the underlying causes of BBB disturbance, which include ageing, insulin resistance, neurological disorders, heart failure and methamphetamine abuse. It is reasonable to conclude that exercise may represent a good strategy for improving BBB integrity and function.

6 | Summary and Future Direction

Exercise confers significant benefits across all ages and sexes (Figure 7), improving brain-related functions such as cognitive performance, mood regulation, sleep quality and mental health conditions like depression and dementia. These effects seem to be mediated by metabolic improvements including enhanced insulin sensitivity, reduced inflammation and cardiovascular health [238], as well as exerkines/myokines like BDNF

(Figure 3), CSF1 [140, 143] and cathepsin B [171]. The influence of exercise on the heart-brain axis is promising and merits deeper exploration [239].

The preventive and therapeutic potential of exercise enhances neuroplasticity and supports recovery, particularly in CNS disorders characterised by demyelination, like multiple sclerosis. The evidence underscores the importance of incorporating exercise into strategies designed to support ageing populations and manage neurodegenerative diseases.

Several questions still remain for future investigation. For instance, how do different types of physical activity affect specific CNS regions? What myokines and exerkines are produced in response to different forms of exercise, and how do these differ by exercise modality? Furthermore, how do male and female CNS and cardiovascular systems respond to exercise, and what are the differential effects of physical activity on the ageing CNS? Answers to these questions will advance our understanding of the mechanisms underlying exercise-induced CNS benefits and refine targeted interventions for diverse populations.

Author Contributions

Conceptualisation, manuscript preparation and revision: S.P., C.A.D. Manuscript review and revision: F.A.N., R.K.M. All authors have read and approved the final version of the manuscript.

Acknowledgements

The Figures are created with [BioRender.com](https://www.bio-render.com/).

The authors used ChatGPT to assist with grammar and clarity during the preparation of this manuscript. All content was subsequently reviewed and edited by the authors, who take full responsibility for the final text.

Conflicts of Interest

The authors declare no conflicts of interest.

Data Availability Statement

All data supporting this study are available in the published literature, as cited in the manuscript.

References

1. B. Dubois, H. H. Feldman, C. Jacova, et al., "Revising the Definition of Alzheimer's Disease: A New Lexicon," *Lancet Neurology* 9 (2010): 1118–1127.
2. Z. Arvanitakis, R. C. Shah, and D. A. Bennett, "Diagnosis and Management of Dementia: Review," *JAMA* 322 (2019): 1589–1599.
3. S. Norton, F. E. Matthews, D. E. Barnes, K. Yaffe, and C. Brayne, "Potential for Primary Prevention of Alzheimer's Disease: An Analysis of Population-Based Data," *Lancet Neurology* 13 (2014): 788–794.
4. P. T. Kamatham, R. Shukla, D. K. Khatri, and L. K. Vora, "Pathogenesis, Diagnostics, and Therapeutics for Alzheimer's Disease: Breaking the Memory Barrier," *Ageing Research Reviews* 101 (2024): 102481.
5. E. Jaul and J. Barron, "Age-Related Diseases and Clinical and Public Health Implications for the 85 Years Old and Over Population," *Frontiers in Public Health* 5 (2017): 335.
6. D. J. Tisserand, P. J. Visser, M. P. van Boxtel, and J. Jolles, "The Relation Between Global and Limbic Brain Volumes on MRI and Cognitive Performance in Healthy Individuals Across the Age Range," *Neurobiology of Aging* 21 (2000): 569–576.
7. D. J. Tisserand, J. C. Pruessner, E. J. Sanz Arigita, et al., "Regional Frontal Cortical Volumes Decrease Differentially in Aging: An MRI Study to Compare Volumetric Approaches and Voxel-Based Morphometry," *NeuroImage* 17 (2002): 657–669.
8. E. R. Sowell, B. S. Peterson, P. M. Thompson, S. E. Welcome, A. L. Henkenius, and A. W. Toga, "Mapping Cortical Change Across the Human Life Span," *Nature Neuroscience* 6 (2003): 309–315.
9. E. R. Sowell, P. M. Thompson, K. D. Tessner, and A. W. Toga, "Mapping Continued Brain Growth and Gray Matter Density Reduction in Dorsal Frontal Cortex: Inverse Relationships During Postadolescent Brain Maturation," *Journal of Neuroscience* 21 (2001): 8819–8829.
10. N. Raz, U. Lindenberger, K. M. Rodrigue, et al., "Regional Brain Changes in Aging Healthy Adults: General Trends, Individual Differences and Modifiers," *Cerebral Cortex (New York, NY: 1991)* 15 (2005): 1676–1689.
11. R. Peters, "Ageing and the Brain," *Postgraduate Medical Journal* 82 (2006): 84–88.
12. T. D. Smith, M. M. Adams, M. Gallagher, J. H. Morrison, and P. R. Rapp, "Circuit-Specific Alterations in Hippocampal Synaptophysin Immunoreactivity Predict Spatial Learning Impairment in Aged Rats," *Journal of Neuroscience* 20 (2000): 6587–6593.
13. C. A. Barnes, "Normal Aging: Regionally Specific Changes in Hippocampal Synaptic Transmission," *Trends in Neurosciences* 17 (1994): 13–18.
14. C. T. Siwak-Tapp, E. Head, B. A. Muggenburg, N. W. Milgram, and C. W. Cotman, "Neurogenesis Decreases With Age in the Canine Hippocampus and Correlates With Cognitive Function," *Neurobiology of Learning and Memory* 88 (2007): 249–259.
15. H. G. Kuhn, H. Dickinson-Anson, and F. H. Gage, "Neurogenesis in the Dentate Gyrus of the Adult Rat: Age-Related Decrease of Neuronal Progenitor Proliferation," *Journal of Neuroscience* 16 (1996): 2027–2033.
16. R. O. Roberts, Y. E. Geda, D. S. Knopman, et al., "Cardiac Disease Associated With Increased Risk of Nonamnesic Cognitive Impairment: Stronger Effect on Women," *JAMA Neurology* 70 (2013): 374–382.
17. K. Deckers, S. H. J. Schievink, M. M. F. Rodriguez, et al., "Coronary Heart Disease and Risk for Cognitive Impairment or Dementia: Systematic Review and Meta-Analysis," *PLoS One* 12 (2017): e0184244.
18. C. A. Hammond, N. J. Blades, S. I. Chaudhry, et al., "Long-Term Cognitive Decline After Newly Diagnosed Heart Failure: Longitudinal Analysis in the CHS (Cardiovascular Health Study)," *Circulation. Heart Failure* 11 (2018): e004476.
19. W. Xie, F. Zheng, L. Yan, and B. Zhong, "Cognitive Decline Before and After Incident Coronary Events," *Journal of the American College of Cardiology* 73 (2019): 3041–3050.
20. D. M. Lloyd-Jones, E. P. Leip, M. G. Larson, et al., "Prediction of Lifetime Risk for Cardiovascular Disease by Risk Factor Burden at 50 Years of Age," *Circulation* 113 (2006): 791–798.
21. J. Agrimi, D. Menicucci, J. H. Qu, et al., "Enhanced Myocardial Adenylate Cyclase Activity Alters Heart-Brain Communication," *JACC: Clinical Electrophysiology* 9 (2023): 2219–2235.
22. J. H. Park, J. H. Moon, H. J. Kim, M. H. Kong, and Y. H. Oh, "Sedentary Lifestyle: Overview of Updated Evidence of Potential Health Risks," *Korean Journal of Family Medicine* 41 (2020): 365–373.
23. M. P. Mattson, "Evolutionary Aspects of Human Exercise—Born to Run Purposefully," *Ageing Research Reviews* 11 (2012): 347–352.
24. J. A. Kanaley, S. R. Colberg, M. H. Corcoran, et al., "Exercise/Physical Activity in Individuals With Type 2 Diabetes: A Consensus Statement From the American College of Sports Medicine," *Medicine and Science in Sports and Exercise* 54 (2022): 353–368.
25. C. Greenhill, "The Metabolic Benefits of Exercise-Induced Hepatic Autophagy," *Nature Reviews. Endocrinology* 19, no. 5 (2023): 254, <https://doi.org/10.1038/s41574-023-00823-6>.
26. Y. Park, D. H. Sinn, K. Kim, and G. Y. Gwak, "Associations of Physical Activity Domains and Muscle Strength Exercise With Non-Alcoholic Fatty Liver Disease: A Nation-Wide Cohort Study," *Scientific Reports* 13 (2023): 4724.
27. M. Idorn and P. Hojman, "Exercise-Dependent Regulation of NK Cells in Cancer Protection," *Trends in Molecular Medicine* 22 (2016): 565–577.
28. J. Alty, M. Farrow, and K. Lawler, "Exercise and Dementia Prevention," *Practical Neurology* 20 (2020): 234–240.
29. J. Agrimi, C. Spalletti, C. Baroni, et al., "Obese Mice Exposed to Psychosocial Stress Display Cardiac and Hippocampal Dysfunction Associated With Local Brain-Derived Neurotrophic Factor Depletion," *eBioMedicine* 47 (2019): 384–401.
30. D. J. O'Gorman, H. K. Karlsson, S. McQuaid, et al., "Exercise Training Increases Insulin-Stimulated Glucose Disposal and GLUT4 (SLC2A4) Protein Content in Patients With Type 2 Diabetes," *Diabetologia* 49 (2006): 2983–2992.
31. A. N. Pizarro, J. C. Ribeiro, E. A. Marques, J. Mota, and M. P. Santos, "Is Walking to School Associated With Improved Metabolic Health?," *International Journal of Behavioral Nutrition and Physical Activity* 10 (2013): 12.
32. T. Christiansen, S. K. Paulsen, J. M. Bruun, S. B. Pedersen, and B. Richelsen, "Exercise Training Versus Diet-Induced Weight-Loss on Metabolic Risk Factors and Inflammatory Markers in Obese Subjects: A 12-Week Randomized Intervention Study," *American Journal of Physiology. Endocrinology and Metabolism* 298 (2010): E824–E831.
33. N. M. Hamburg, C. J. McMackin, A. L. Huang, et al., "Physical Inactivity Rapidly Induces Insulin Resistance and Microvascular Dysfunction in Healthy Volunteers," *Arteriosclerosis, Thrombosis, and Vascular Biology* 27 (2007): 2650–2656.
34. G. N. Healy, D. W. Dunstan, J. Salmon, et al., "Breaks in Sedentary Time: Beneficial Associations With Metabolic Risk," *Diabetes Care* 31 (2008): 661–666.

35. D. W. Dunstan, B. A. Kingwell, R. Larsen, et al., "Breaking Up Prolonged Sitting Reduces Postprandial Glucose and Insulin Responses," *Diabetes Care* 35 (2012): 976–983.
36. J. Kusuyama, A. B. Alves-Wagner, N. S. Makarewicz, and L. J. Goodyear, "Effects of Maternal and Paternal Exercise on Offspring Metabolism," *Nature Metabolism* 2 (2020): 858–872.
37. R. C. Laker, A. Altintas, T. S. Lillard, et al., "Exercise During Pregnancy Mitigates Negative Effects of Parental Obesity on Metabolic Function in Adult Mouse Offspring," *Journal of Applied Physiology* (Bethesda, MD: 1985) 130 (2021): 605–616.
38. C. Moyer, O. R. Reoyo, and L. May, "The Influence of Prenatal Exercise on Offspring Health: A Review," *Clinical Medicine Insights: Women's Health* 9 (2016): 37–42.
39. K. K. Garnaes, S. A. Nyrenes, K. A. Salvesen, O. Salvesen, S. Morkved, and T. Moholdt, "Effect of Supervised Exercise Training During Pregnancy on Neonatal and Maternal Outcomes Among Overweight and Obese Women. Secondary Analyses of the ETIP Trial: A Randomised Controlled Trial," *PLoS One* 12 (2017): e0173937.
40. J. F. Clapp, B. Lopez, and R. Harcar-Sevcik, "Neonatal Behavioral Profile of the Offspring of Women Who Continued to Exercise Regularly Throughout Pregnancy," *American Journal of Obstetrics and Gynecology* 180 (1999): 91–94.
41. L. E. May, S. A. Scholtz, R. Suminski, and K. M. Gustafson, "Aerobic Exercise During Pregnancy Influences Infant Heart Rate Variability at One Month of Age," *Early Human Development* 90 (2014): 33–38.
42. A. G. McMillan, L. E. May, G. G. Gaines, C. Isler, and D. Kuehn, "Effects of Aerobic Exercise During Pregnancy on 1-Month Infant Neuromotor Skills," *Medicine and Science in Sports and Exercise* 51 (2019): 1671–1676.
43. T. Moholdt and K. I. Stanford, "Exercised Breastmilk: A Kick-Start to Prevent Childhood Obesity?," *Trends in Endocrinology and Metabolism* 35 (2024): 23–30.
44. F. Sofi, D. Valecchi, D. Bacci, et al., "Physical Activity and Risk of Cognitive Decline: A Meta-Analysis of Prospective Studies," *Journal of Internal Medicine* 269 (2011): 107–117.
45. M. Hamer and Y. Chida, "Physical Activity and Risk of Neurodegenerative Disease: A Systematic Review of Prospective Evidence," *Psychological Medicine* 39 (2009): 3–11.
46. D. A. Raichlen and J. D. Polk, "Linking Brains and Brawn: Exercise and the Evolution of Human Neurobiology," *Proceedings of the Biological Sciences* 280 (2013): 2012250.
47. D. A. Raichlen and A. D. Gordon, "Relationship Between Exercise Capacity and Brain Size in Mammals," *PLoS One* 6 (2011): e20601.
48. I. Matthews, A. Birnbaum, A. Gromova, et al., "Skeletal Muscle TFEB Signaling Promotes Central Nervous System Function and Reduces Neuroinflammation During Aging and Neurodegenerative Disease," *Cell Reports* 42 (2023): 113436.
49. F. Herold, A. Torpel, L. Schega, and N. G. Muller, "Functional and/or Structural Brain Changes in Response to Resistance Exercises and Resistance Training Lead to Cognitive Improvements – A Systematic Review," *European Review of Aging and Physical Activity* 16 (2019): 10.
50. K. I. Erickson, R. L. Leckie, and A. M. Weinstein, "Physical Activity, Fitness, and Gray Matter Volume," *Neurobiology of Aging* 35, no. Suppl 2 (2014): S20–S28.
51. N. D. Koblinsky, L.-A. C. Meusel, C. E. Greenwood, and N. D. Anderson, "Household Physical Activity Is Positively Associated With Gray Matter Volume in Older Adults," *BMC Geriatrics* 21 (2021): 104.
52. W. D. S. Killgore, E. A. Olson, and M. Weber, "Physical Exercise Habits Correlate With Gray Matter Volume of the Hippocampus in Healthy Adult Humans," *Scientific Reports* 3 (2013): 3457.
53. K. Wittfeld, C. Jochem, M. Dorr, et al., "Cardiorespiratory Fitness and Gray Matter Volume in the Temporal, Frontal, and Cerebellar Regions in the General Population," *Mayo Clinic Proceedings* 95 (2020): 44–56.
54. S. Colcombe and A. F. Kramer, "Fitness Effects on the Cognitive Function of Older Adults: A Meta-Analytic Study," *Psychological Science* 14 (2003): 125–130.
55. P. J. Smith, J. A. Blumenthal, B. M. Hoffman, et al., "Aerobic Exercise and Neurocognitive Performance: A Meta-Analytic Review of Randomized Controlled Trials," *Psychosomatic Medicine* 72 (2010): 239–252.
56. G. Skovbjerg, A. M. Fritzen, C. S. A. Svendsen, et al., "Atlas of Exercise-Induced Brain Activation in Mice," *Molecular Metabolism* 82 (2024): 101907.
57. A. Montagne, S. R. Barnes, M. D. Sweeney, et al., "Blood-Brain Barrier Breakdown in the Aging Human Hippocampus," *Neuron* 85 (2015): 296–302.
58. J. L. Whitwell, H. J. Wiste, S. D. Weigand, et al., "Comparison of Imaging Biomarkers in the Alzheimer Disease Neuroimaging Initiative and the Mayo Clinic Study of Aging," *Archives of Neurology* 69 (2012): 614–622.
59. L. G. Apostolova, K. S. Hwang, J. P. Andrawis, et al., "3D PIB and CSF Biomarker Associations With Hippocampal Atrophy in ADNI Subjects," *Neurobiology of Aging* 31 (2010): 1284–1303.
60. J. R. Epp, C. Chow, and L. A. Galea, "Hippocampus-Dependent Learning Influences Hippocampal Neurogenesis," *Frontiers in Neuroscience* 7 (2013): 57.
61. H. van Praag, T. Shubert, C. Zhao, and F. H. Gage, "Exercise Enhances Learning and Hippocampal Neurogenesis in Aged Mice," *Journal of Neuroscience* 25 (2005): 8680–8685.
62. D. J. Creer, C. Romberg, L. M. Saksida, H. van Praag, and T. J. Bussey, "Running Enhances Spatial Pattern Separation in Mice," *Proceedings of the National Academy of Sciences of the United States of America* 107 (2010): 2367–2372.
63. M. W. Marlatt, M. C. Potter, P. J. Lucassen, and H. van Praag, "Running Throughout Middle-Age Improves Memory Function, Hippocampal Neurogenesis, and BDNF Levels in Female C57BL/6J Mice," *Developmental Neurobiology* 72 (2012): 943–952.
64. J. E. Fardell, J. Vardy, J. D. Shah, and I. N. Johnston, "Cognitive Impairments Caused by Oxaliplatin and 5-Fluorouracil Chemotherapy Are Ameliorated by Physical Activity," *Psychopharmacology* 220 (2012): 183–193.
65. T. J. Fischer, T. L. Walker, R. W. Overall, M. D. Brandt, and G. Kempermann, "Acute Effects of Wheel Running on Adult Hippocampal Precursor Cells in Mice Are Not Caused by Changes in Cell Cycle Length or S Phase Length," *Frontiers in Neuroscience* 8 (2014): 314.
66. M. C. Lee, K. Inoue, M. Okamoto, et al., "Voluntary Resistance Running Induces Increased Hippocampal Neurogenesis in Rats Comparable to Load-Free Running," *Neuroscience Letters* 537 (2013): 6–10.
67. E. Castilla-Ortega, C. Rosell-Valle, C. Pedraza, F. de Rodriguez Fonseca, G. Estivill-Torrus, and L. J. Santin, "Voluntary Exercise Followed by Chronic Stress Strikingly Increases Mature Adult-Born Hippocampal Neurons and Prevents Stress-Induced Deficits in 'What-When-Where' Memory," *Neurobiology of Learning and Memory* 109 (2014): 62–73.
68. J. C. Conover and R. L. Allen, "The Subventricular Zone: New Molecular and Cellular Developments," *Cellular and Molecular Life Sciences* 59 (2002): 2128–2135.
69. A. Castells-Sanchez, F. Roig-Coll, R. Dacosta-Aguayo, et al., "Exercise and Fitness Neuroprotective Effects: Molecular, Brain Volume and Psychological Correlates and Their Mediating Role in Healthy Late-Middle-Aged Women and Men," *Frontiers in Aging Neuroscience* 13 (2021): 615247.

70. H. Yanagisawa, I. Dan, D. Tsuzuki, et al., "Acute Moderate Exercise Elicits Increased Dorsolateral Prefrontal Activation and Improves Cognitive Performance With Stroop Test," *NeuroImage* 50 (2010): 1702–1710.
71. S. J. Colcombe, K. I. Erickson, N. Raz, et al., "Aerobic Fitness Reduces Brain Tissue Loss in Aging Humans," *Journals of Gerontology. Series A, Biological Sciences and Medical Sciences* 58 (2003): 176–180.
72. A. C. Smolarek, L. H. Ferreira, L. P. Mascarenhas, et al., "The Effects of Strength Training on Cognitive Performance in Elderly Women," *Clinical Interventions in Aging* 11 (2016): 749–754.
73. K. I. Erickson, M. W. Voss, R. S. Prakash, et al., "Exercise Training Increases Size of Hippocampus and Improves Memory," *Proceedings of the National Academy of Sciences of the United States of America* 108 (2011): 3017–3022.
74. L. S. Nagamatsu, T. C. Handy, C. L. Hsu, M. Voss, and T. Liu-Ambrose, "Resistance Training Promotes Cognitive and Functional Brain Plasticity in Seniors With Probable Mild Cognitive Impairment," *Archives of Internal Medicine* 172 (2012): 666–668.
75. D. H. Yoon, D. Kang, H. J. Kim, J. S. Kim, H. S. Song, and W. Song, "Effect of Elastic Band-Based High-Speed Power Training on Cognitive Function, Physical Performance and Muscle Strength in Older Women With Mild Cognitive Impairment," *Geriatrics & Gerontology International* 17 (2017): 765–772.
76. L. S. Chow, R. E. Gerszten, J. M. Taylor, et al., "Exerkines in Health, Resilience and Disease," *Nature Reviews. Endocrinology* 18 (2022): 273–289.
77. A. I. Doncheva, S. Romero, M. Ramirez-Garrastacho, et al., "Extracellular Vesicles and microRNAs Are Altered in Response to Exercise, Insulin Sensitivity and Overweight," *Acta Physiologica (Oxford, England)* 236 (2022): e13862.
78. C. Morland, K. A. Andersson, O. P. Haugen, et al., "Exercise Induces Cerebral VEGF and Angiogenesis via the Lactate Receptor HCAR1," *Nature Communications* 8 (2017): 15557.
79. M. Rai and F. Demontis, "Muscle-to-Brain Signaling via Myokines and Myometabolites," *Brain Plasticity* 8 (2022): 43–63.
80. D. Garcia-Rodriguez and A. Gimenez-Cassina, "Ketone Bodies in the Brain Beyond Fuel Metabolism: From Excitability to Gene Expression and Cell Signaling," *Frontiers in Molecular Neuroscience* 14 (2021): 732120.
81. R. Yao, K. Nishii, N. Aizu, T. Kito, K. Sakai, and K. Yamada, "Maintenance of the Amygdala-Hippocampal Circuit Function With Safe and Feasible Shaking Exercise Therapy in SAMP-10 Mice," *Dementia and Geriatric Cognitive Disorders Extra* 11 (2021): 114–121.
82. T. Tsujii, K. Komatsu, and K. Sakatani, "Acute Effects of Physical Exercise on Prefrontal Cortex Activity in Older Adults: A Functional Near-Infrared Spectroscopy Study," *Advances in Experimental Medicine and Biology* 765 (2013): 293–298.
83. R. Lima-Filho, J. S. Fortuna, D. Cozachenko, et al., "Brain FNDC5/Irisin Expression in Patients and Mouse Models of Major Depression," *eNeuro* 10, no. 2 (2023): ENEURO.0256-22.2023, <https://doi.org/10.1523/ENEURO.0256-22.2023>.
84. E. Albrecht, L. Schering, F. Buck, et al., "Irisin: Still Chasing Shadows," *Molecular Metabolism* 34 (2020): 124–135.
85. S. Maak, F. Norheim, C. A. Drevon, and H. P. Erickson, "Progress and Challenges in the Biology of FNDC5 and Irisin," *Endocrine Reviews* 42 (2021): 436–456.
86. A. M. Horowitz, X. Fan, G. Bieri, et al., "Blood Factors Transfer Beneficial Effects of Exercise on Neurogenesis and Cognition to the Aged Brain," *Science* 369 (2020): 167–173.
87. Z. De Miguel, N. Khoury, M. J. Betley, et al., "Exercise Plasma Boosts Memory and Dampens Brain Inflammation via Clusterin," *Nature* 600 (2021): 494–499.
88. A. C. McPherron, A. M. Lawler, and S. J. Lee, "Regulation of Skeletal Muscle Mass in Mice by a New TGF-Beta Superfamily Member," *Nature* 387 (1997): 83–90.
89. B. K. Pedersen, A. Steensberg, C. Fischer, et al., "Searching for the Exercise Factor: Is IL-6 a Candidate?," *Journal of Muscle Research and Cell Motility* 24 (2003): 113–119.
90. P. Bostrom, J. Wu, M. P. Jedrychowski, et al., "A PGC1-Alpha-Dependent Myokine That Drives Brown-Fat-Like Development of White Fat and Thermogenesis," *Nature* 481 (2012): 463–468.
91. K. Sakuma, K. Watanabe, M. Sano, et al., "A Possible Role for BDNF, NT-4 and TrkB in the Spinal Cord and Muscle of Rat Subjected to Mechanical Overload, Bupivacaine Injection and Axotomy," *Brain Research* 907 (2001): 1–19.
92. S. Edman, O. Horwath, T. Van der Stede, et al., "Pro-Brain-Derived Neurotrophic Factor (BDNF), but Not Mature BDNF, Is Expressed in Human Skeletal Muscle: Implications for Exercise-Induced Neuroplasticity," *Function (Oxford, England)* 5 (2024): zqae005.
93. T. Usui, A. Naruo, M. Okada, Y. Hayabe, and H. Yamawaki, "Brain-Derived Neurotrophic Factor Promotes Angiogenic Tube Formation Through Generation of Oxidative Stress in Human Vascular Endothelial Cells," *Acta Physiologica (Oxford, England)* 211 (2014): 385–394.
94. P. Kermani and B. Hempstead, "Brain-Derived Neurotrophic Factor: A Newly Described Mediator of Angiogenesis," *Trends in Cardiovascular Medicine* 17 (2007): 140–143.
95. M. M. Hofer and Y. A. Barde, "Brain-Derived Neurotrophic Factor Prevents Neuronal Death In Vivo," *Nature* 331 (1988): 261–262.
96. M. P. Mattson, S. Maudsley, and B. Martin, "BDNF and 5-HT: A Dynamic Duo in Age-Related Neuronal Plasticity and Neurodegenerative Disorders," *Trends in Neurosciences* 27 (2004): 589–594.
97. M. E. Greenberg, B. Xu, B. Lu, and B. L. Hempstead, "New Insights in the Biology of BDNF Synthesis and Release: Implications in CNS Function," *Journal of Neuroscience* 29 (2009): 12764–12767.
98. S. J. Mowla, S. Pareek, H. F. Farhadi, et al., "Differential Sorting of Nerve Growth Factor and Brain-Derived Neurotrophic Factor in Hippocampal Neurons," *Journal of Neuroscience* 19 (1999): 2069–2080.
99. T. Mizui, Y. Ishikawa, H. Kumanogoh, et al., "BDNF Pro-Peptide Actions Facilitate Hippocampal LTD and Are Altered by the Common BDNF Polymorphism Val66Met," *Proceedings of the National Academy of Sciences of the United States of America* 112 (2015): E3067–E3074.
100. J. P. Zanin, N. Unsain, and A. Anastasia, "Growth Factors and Hormones Pro-Peptides: The Unexpected Adventures of the BDNF Prodomain," *Journal of Neurochemistry* 141 (2017): 330–340.
101. J. M. Conner, J. C. Lauterborn, Q. Yan, C. M. Gall, and S. Varon, "Distribution of Brain-Derived Neurotrophic Factor (BDNF) Protein and mRNA in the Normal Adult Rat CNS: Evidence for Anterograde Axonal Transport," *Journal of Neuroscience* 17 (1997): 2295–2313.
102. A. Luoni, F. Macchi, M. Papp, R. Molteni, and M. A. Riva, "Lurasidone Exerts Antidepressant Properties in the Chronic Mild Stress Model Through the Regulation of Synaptic and Neuroplastic Mechanisms in the Rat Prefrontal Cortex," *International Journal of Neuropsychopharmacology* 18, no. 4 (2014): pyu061, <https://doi.org/10.1093/ijnp/pyu061>.
103. P. Kowianski, G. Lietzau, E. Czuba, M. Waskow, A. Steliga, and J. Morys, "BDNF: A Key Factor With Multipotent Impact on Brain Signaling and Synaptic Plasticity," *Cellular and Molecular Neurobiology* 38 (2018): 579–593.
104. G. Fulgenzi, F. Tomassoni-Ardori, L. Babini, et al., "BDNF Modulates Heart Contraction Force and Long-Term Homeostasis Through Truncated TrkB.T1 Receptor Activation," *Journal of Cell Biology* 210 (2015): 1003–1012.

105. K. Mousavi and B. J. Jasmin, "BDNF Is Expressed in Skeletal Muscle Satellite Cells and Inhibits Myogenic Differentiation," *Journal of Neuroscience* 26 (2006): 5739–5749.
106. C. Cunha, R. Brambilla, and K. L. Thomas, "A Simple Role for BDNF in Learning and Memory?," *Frontiers in Molecular Neuroscience* 3 (2010): 1.
107. P. Bekinschtein, M. Cammarota, C. Katche, et al., "BDNF Is Essential to Promote Persistence of Long-Term Memory Storage," *Proceedings of the National Academy of Sciences of the United States of America* 105 (2008): 2711–2716.
108. P. Bekinschtein, M. Cammarota, and J. H. Medina, "BDNF and Memory Processing," *Neuropharmacology* 76 (2014): 677–683.
109. P. Bekinschtein, M. Cammarota, I. Izquierdo, and J. H. Medina, "BDNF and Memory Formation and Storage," *Neuroscientist* 14 (2008): 147–156.
110. F. Cirulli, A. Berry, F. Chiarotti, and E. Alleva, "Intrahippocampal Administration of BDNF in Adult Rats Affects Short-Term Behavioral Plasticity in the Morris Water Maze and Performance in the Elevated Plus-Maze," *Hippocampus* 14 (2004): 802–807.
111. J. S. Mu, W. P. Li, Z. B. Yao, and X. F. Zhou, "Deprivation of Endogenous Brain-Derived Neurotrophic Factor Results in Impairment of Spatial Learning and Memory in Adult Rats," *Brain Research* 835 (1999): 259–265.
112. H. W. Horch, A. Kruttgen, S. D. Portbury, and L. C. Katz, "Destabilization of Cortical Dendrites and Spines by BDNF," *Neuron* 23 (1999): 353–364.
113. L. T. Ferris, J. S. Williams, and C. L. Shen, "The Effect of Acute Exercise on Serum Brain-Derived Neurotrophic Factor Levels and Cognitive Function," *Medicine and Science in Sports and Exercise* 39 (2007): 728–734.
114. E. W. Griffin, S. Mullally, C. Foley, S. A. Warmington, S. M. O'Mara, and A. M. Kelly, "Aerobic Exercise Improves Hippocampal Function and Increases BDNF in the Serum of Young Adult Males," *Physiology & Behavior* 104 (2011): 934–941.
115. P. Rasmussen, P. Brassard, H. Adser, et al., "Evidence for a Release of Brain-Derived Neurotrophic Factor From the Brain During Exercise," *Experimental Physiology* 94 (2009): 1062–1069.
116. S. W. Tang, E. Chu, T. Hui, D. Helmeste, and C. Law, "Influence of Exercise on Serum Brain-Derived Neurotrophic Factor Concentrations in Healthy Human Subjects," *Neuroscience Letters* 431 (2008): 62–65.
117. S. Rojas Vega, H. K. Struder, B. Vera Wahrmann, A. Schmidt, W. Bloch, and W. Hollmann, "Acute BDNF and Cortisol Response to Low Intensity Exercise and Following Ramp Incremental Exercise to Exhaustion in Humans," *Brain Research* 1121 (2006): 59–65.
118. J. A. Zoladz, A. Pilc, J. Majerczak, M. Grandys, J. Zapart-Bukowska, and K. Duda, "Endurance Training Increases Plasma Brain-Derived Neurotrophic Factor Concentration in Young Healthy Men," *Journal of Physiology and Pharmacology* 59, no. Suppl 7 (2008): 119–132.
119. J. M. Gaitan, H. Y. Moon, M. Stremlau, et al., "Effects of Aerobic Exercise Training on Systemic Biomarkers and Cognition in Late Middle-Aged Adults at Risk for Alzheimer's Disease," *Frontiers in Endocrinology (Lausanne)* 12 (2021): 660181.
120. D. Kimhy, J. Vakhrusheva, M. N. Bartels, et al., "The Impact of Aerobic Exercise on Brain-Derived Neurotrophic Factor and Neurocognition in Individuals With Schizophrenia: A Single-Blind, Randomized Clinical Trial," *Schizophrenia Bulletin* 41 (2015): 859–868.
121. J. A. Zoladz, J. Majerczak, E. Zeligowska, et al., "Moderate-Intensity Interval Training Increases Serum Brain-Derived Neurotrophic Factor Level and Decreases Inflammation in Parkinson's Disease Patients," *Journal of Physiology and Pharmacology* 65 (2014): 441–448.
122. A. V. Araya, X. Orellana, D. Godoy, L. Soto, and J. Fiedler, "Effect of Exercise on Circulating Levels of Brain-Derived Neurotrophic Factor (BDNF) in Overweight and Obese Subjects," *Hormone and Metabolic Research* 45 (2013): 541–544.
123. J. K. Lee, A. C. Koh, S. X. Koh, G. J. Liu, A. Q. Nio, and P. W. Fan, "Neck Cooling and Cognitive Performance Following Exercise-Induced Hyperthermia," *European Journal of Applied Physiology* 114 (2014): 375–384.
124. K. Skriver, M. Roig, J. Lundbye-Jensen, et al., "Acute Exercise Improves Motor Memory: Exploring Potential Biomarkers," *Neurobiology of Learning and Memory* 116 (2014): 46–58.
125. B. Winter, C. Breitenstein, F. C. Mooren, et al., "High Impact Running Improves Learning," *Neurobiology of Learning and Memory* 87 (2007): 597–609.
126. J. J. Heisz, I. B. Clark, K. Bonin, et al., "The Effects of Physical Exercise and Cognitive Training on Memory and Neurotrophic Factors," *Journal of Cognitive Neuroscience* 29 (2017): 1895–1907.
127. P. Z. Liu and R. Nusslock, "Exercise-Mediated Neurogenesis in the Hippocampus via BDNF," *Frontiers in Neuroscience* 12 (2018): 52.
128. M. Parrini, D. Ghezzi, G. Deidda, et al., "Aerobic Exercise and a BDNF-Mimetic Therapy Rescue Learning and Memory in a Mouse Model of Down Syndrome," *Scientific Reports* 7 (2017): 16825.
129. W. Pan, W. A. Banks, M. B. Fasold, J. Bluth, and A. J. Kastin, "Transport of Brain-Derived Neurotrophic Factor Across the Blood-Brain Barrier," *Neuropharmacology* 37 (1998): 1553–1561.
130. J. F. Yarrow, L. J. White, S. C. McCoy, and S. E. Borst, "Training Augments Resistance Exercise Induced Elevation of Circulating Brain Derived Neurotrophic Factor (BDNF)," *Neuroscience Letters* 479 (2010): 161–165.
131. F. Norheim, T. Raastad, B. Thiede, A. C. Rustan, C. A. Drevon, and F. Haugen, "Proteomic Identification of Secreted Proteins From Human Skeletal Muscle Cells and Expression in Response to Strength Training," *American Journal of Physiology. Endocrinology and Metabolism* 301 (2011): E1013–E1021.
132. A. Cannavo, S. Jun, G. Rengo, et al., "beta3AR-Dependent Brain-Derived Neurotrophic Factor (BDNF) Generation Limits Chronic Postischemic Heart Failure," *Circulation Research* 132 (2023): 867–881.
133. N. Feng, S. Huke, G. Zhu, et al., "Constitutive BDNF/TrkB Signaling Is Required for Normal Cardiac Contraction and Relaxation," *Proceedings of the National Academy of Sciences of the United States of America* 112 (2015): 1880–1885.
134. X. Yang, M. Zhang, B. Xie, et al., "Myocardial Brain-Derived Neurotrophic Factor Regulates Cardiac Bioenergetics Through the Transcription Factor Yin Yang 1," *Cardiovascular Research* 119 (2023): 571–586.
135. A. Elia, A. Cannavo, G. Gambino, et al., "Aging Is Associated With Cardiac Autonomic Nerve Fiber Depletion and Reduced Cardiac and Circulating BDNF Levels," *Journal of Geriatric Cardiology* 18 (2021): 549–559.
136. S. Pourteymour, K. Eckardt, T. Holen, et al., "Global mRNA Sequencing of Human Skeletal Muscle: Search for Novel Exercise-Regulated Myokines," *Molecular Metabolism* 6, no. 4 (2017): 352–365, <https://doi.org/10.1016/j.molmet.2017.01.007>.
137. T. M. Langleite, J. Jensen, F. Norheim, et al., "Insulin Sensitivity, Body Composition and Adipose Depots Following 12 w Combined Endurance and Strength Training in Dysglycemic and Normoglycemic Sedentary Men," *Archives of Physiology and Biochemistry* 122 (2016): 167–179.
138. C. Easley-Neal, O. Foreman, N. Sharma, A. A. Zarrin, and R. M. Weimer, "CSF1R Ligands IL-34 and CSF1 Are Differentially Required for Microglia Development and Maintenance in White and Gray Matter Brain Regions," *Frontiers in Immunology* 10 (2019): 2199.

139. J. A. Hamilton, "Colony-Stimulating Factors in Inflammation and Autoimmunity," *Nature Reviews. Immunology* 8 (2008): 533–544.
140. N. Oosterhof, L. E. Kuil, H. C. van der Linde, et al., "Colony-Stimulating Factor 1 Receptor (CSF1R) Regulates Microglia Density and Distribution, but Not Microglia Differentiation In Vivo," *Cell Reports* 24 (2018): 1203–1217.e1206.
141. R. Berglund, Y. Cheng, E. Piket, et al., "The Aging Mouse CNS Is Protected by an Autophagy-Dependent Microglia Population Promoted by IL-34," *Nature Communications* 15 (2024): 383.
142. E. R. Stanley and P. M. Heard, "Factors Regulating Macrophage Production and Growth. Purification and Some Properties of the Colony Stimulating Factor From Medium Conditioned by Mouse L Cells," *Journal of Biological Chemistry* 252 (1977): 4305–4312.
143. H. Lin, E. Lee, K. Hestir, et al., "Discovery of a Cytokine and Its Receptor by Functional Screening of the Extracellular Proteome," *Science* 320 (2008): 807–811.
144. S. J. Hwang, B. Choi, S. S. Kang, et al., "Interleukin-34 Produced by Human Fibroblast-Like Synovial Cells in Rheumatoid Arthritis Supports Osteoclastogenesis," *Arthritis Research & Therapy* 14 (2012): R14.
145. Y. Tian, H. Shen, L. Xia, and J. Lu, "Elevated Serum and Synovial Fluid Levels of Interleukin-34 in Rheumatoid Arthritis: Possible Association With Disease Progression via Interleukin-17 Production," *Journal of Interferon & Cytokine Research* 33 (2013): 398–401.
146. B. Rich, M. Scadeng, M. Yamaguchi, P. D. Wagner, and E. C. Breen, "Skeletal Myofiber Vascular Endothelial Growth Factor Is Required for the Exercise Training-Induced Increase in Dentate Gyrus Neuronal Precursor Cells," *Journal of Physiology* 595 (2017): 5931–5943.
147. L. Jensen, H. Pilegaard, P. D. Neuffer, and Y. Hellsten, "Effect of Acute Exercise and Exercise Training on VEGF Splice Variants in Human Skeletal Muscle," *American Journal of Physiology. Regulatory, Integrative and Comparative Physiology* 287 (2004): R397–R402.
148. C. Brinkmann, L. Schafer, M. Masoud, et al., "Effects of Cycling and Exergaming on Neurotrophic Factors in Elderly Type 2 Diabetic Men – A Preliminary Investigation," *Experimental and Clinical Endocrinology & Diabetes* 125 (2017): 436–440.
149. F. Suhr, S. Knuth, S. Achtzehn, J. Mester, and M. de Marees, "Acute Exhaustive Exercise Under Normoxic and Normobaric Hypoxic Conditions Differentially Regulates Angiogenic Biomarkers in Humans," *Medicina (Kaunas, Lithuania)* 57, no. 7 (2021): 727, <https://doi.org/10.3390/medicina57070727>.
150. K. Fabel, K. Fabel, B. Tam, et al., "VEGF Is Necessary for Exercise-Induced Adult Hippocampal Neurogenesis," *European Journal of Neuroscience* 18 (2003): 2803–2812.
151. S. Zhong, L. Li, Y. L. Zhang, et al., "Acetaldehyde Dehydrogenase 2 Interactions With LDLR and AMPK Regulate Foam Cell Formation," *Journal of Clinical Investigation* 129 (2019): 252–267.
152. C. Vivar, M. C. Potter, and H. van Praag, "All About Running: Synaptic Plasticity, Growth Factors and Adult Hippocampal Neurogenesis," *Current Topics in Behavioral Neurosciences* 15 (2013): 189–210.
153. V. Mohamed-Ali, S. Goodrick, A. Rawesh, et al., "Subcutaneous Adipose Tissue Releases Interleukin-6, but Not Tumor Necrosis Factor-Alpha, In Vivo," *Journal of Clinical Endocrinology and Metabolism* 82 (1997): 4196–4200.
154. B. K. Pedersen and M. A. Febbraio, "Muscle as an Endocrine Organ: Focus on Muscle-Derived Interleukin-6," *Physiological Reviews* 88 (2008): 1379–1406.
155. A. Steensberg, G. van Hall, T. Osada, M. Sacchetti, B. Saltin, and B. Klarlund Pedersen, "Production of Interleukin-6 in Contracting Human Skeletal Muscles Can Account for the Exercise-Induced Increase in Plasma Interleukin-6," *Journal of Physiology* 529, no. Pt 1 (2000): 237–242.
156. A. Vilotic, M. Nacka-Aleksic, A. Pirkovic, Z. Bojic-Trbojevic, D. Dekanski, and M. Jovanovic Krivokuca, "IL-6 and IL-8: An Overview of Their Roles in Healthy and Pathological Pregnancies," *International Journal of Molecular Sciences* 23, no. 23 (2022): 14574, <https://doi.org/10.3390/ijms232314574>.
157. B. Schöbitz, D. A. M. Voorhuis, and E. R. De Kloet, "Localization of Interleukin 6 mRNA and Interleukin 6 Receptor mRNA in Rat Brain," *Neuroscience Letters* 136 (1992): 189–192.
158. B. Schöbitz, E. R. de Kloet, W. Sutanto, and F. Holsboer, "Cellular Localization of Interleukin 6 mRNA and Interleukin 6 Receptor mRNA in Rat Brain," *European Journal of Neuroscience* 5 (1993): 1426–1435.
159. R. A. Gadiant and U. Otten, "Differential Expression of Interleukin-6 (IL-6) and Interleukin-6 Receptor (IL-6R) mRNAs in Rat Hypothalamus," *Neuroscience Letters* 153 (1993): 13–16.
160. R. A. Gadiant and U. Otten, "Identification of Interleukin-6 (IL-6)-Expressing Neurons in the Cerebellum and Hippocampus of Normal Adult Rats," *Neuroscience Letters* 182 (1994): 243–246.
161. R. A. Gadiant and U. Otten, "Expression of Interleukin-6 (IL-6) and Interleukin-6 Receptor (IL-6R) mRNAs in Rat Brain During Postnatal Development," *Brain Research* 637 (1994): 10–14.
162. E. Y. Ting, A. C. Yang, and S. J. Tsai, "Role of Interleukin-6 in Depressive Disorder," *International Journal of Molecular Sciences* 21, no. 6 (2020): 2194, <https://doi.org/10.3390/ijms21062194>.
163. E. Haroon, A. W. Daguanno, B. J. Woolwine, et al., "Antidepressant Treatment Resistance Is Associated With Increased Inflammatory Markers in Patients With Major Depressive Disorder," *Psychoneuroendocrinology* 95 (2018): 43–49.
164. M. M. Borovcanin, I. Jovanovic, G. Radosavljevic, et al., "Interleukin-6 in Schizophrenia-Is There a Therapeutic Relevance?," *Frontiers in Psychiatry* 8 (2017): 221.
165. D. R. Goldsmith, E. Haroon, A. H. Miller, G. P. Strauss, P. F. Buckley, and B. J. Miller, "TNF-Alpha and IL-6 Are Associated With the Deficit Syndrome and Negative Symptoms in Patients With Chronic Schizophrenia," *Schizophrenia Research* 199 (2018): 281–284.
166. S. Hidese, J. Matsuo, I. Ishida, et al., "Relationship of Handgrip Strength and Body Mass Index With Cognitive Function in Patients With Schizophrenia," *Frontiers in Psychiatry* 9 (2018): 156.
167. F. Norheim, Y. Hasin-Brumshtein, L. Vergnes, et al., "Gene-by-Sex Interactions in Mitochondrial Functions and Cardio-Metabolic Traits," *Cell Metabolism* 29 (2019): 932–949.e4.
168. H. Y. Moon, A. Becke, D. Berron, et al., "Running-Induced Systemic Cathepsin B Secretion Is Associated With Memory Function," *Cell Metabolism* 24 (2016): 332–340.
169. A. de la Rosa, E. Solana, R. Corpas, et al., "Long-Term Exercise Training Improves Memory in Middle-Aged Men and Modulates Peripheral Levels of BDNF and Cathepsin B," *Scientific Reports* 9 (2019): 3337.
170. B. K. Pedersen, "Physical Activity and Muscle-Brain Crosstalk," *Nature Reviews. Endocrinology* 15 (2019): 383–392.
171. S. Mueller-Stainer, Y. Zhou, H. Arai, et al., "Antiamyloidogenic and Neuroprotective Functions of Cathepsin B: Implications for Alzheimer's Disease," *Neuron* 51 (2006): 703–714.
172. G. Hook, T. Reinheckel, J. Ni, et al., "Cathepsin B Gene Knockout Improves Behavioral Deficits and Reduces Pathology in Models of Neurologic Disorders," *Pharmacological Reviews* 74 (2022): 600–629.
173. Y. Wu, P. Mumford, S. Noy, et al., "Cathepsin B Abundance, Activity and Microglial Localisation in Alzheimer's Disease-Down Syndrome and Early Onset Alzheimer's Disease; the Role of Elevated Cystatin B," *Acta Neuropathologica Communications* 11 (2023): 132.
174. M. E. Raichle and D. A. Gusnard, "Appraising the Brain's Energy Budget," *Proceedings of the National Academy of Sciences of the United States of America* 99 (2002): 10237–10239.

175. Q. Ding, S. Vaynman, P. Souda, J. P. Whitelegge, and F. Gomez-Pinilla, "Exercise Affects Energy Metabolism and Neural Plasticity-Related Proteins in the Hippocampus as Revealed by Proteomic Analysis," *European Journal of Neuroscience* 24 (2006): 1265–1276.
176. N. Ferrara, "Vascular Endothelial Growth Factor and the Regulation of Angiogenesis," *Recent Progress in Hormone Research* 55 (2000): 15–35; discussion 35–16.
177. P. De Rossi, E. Harde, J. P. Dupuis, et al., "A Critical Role for VEGF and VEGFR2 in NMDA Receptor Synaptic Function and Fear-Related Behavior," *Molecular Psychiatry* 21 (2016): 1768–1780.
178. Y. Poitelon, A. M. Kopec, and S. Belin, "Myelin Fat Facts: An Overview of Lipids and Fatty Acid Metabolism," *Cells* 9, no. 4 (2020): 812, <https://doi.org/10.3390/cells9040812>.
179. R. A. Demel and B. De Kruffy, "The Function of Sterols in Membranes," *Biochimica et Biophysica Acta* 457 (1976): 109–132.
180. G. Saher, B. Brugger, C. Lappe-Siefke, et al., "High Cholesterol Level Is Essential for Myelin Membrane Growth," *Nature Neuroscience* 8 (2005): 468–475.
181. N. Camargo, A. Goudriaan, A. F. van Deijk, et al., "Oligodendroglial Myelination Requires Astrocyte-Derived Lipids," *PLoS Biology* 15 (2017): e1002605.
182. J. M. Dietschy and S. D. Turley, "Thematic Review Series: Brain Lipids. Cholesterol Metabolism in the Central Nervous System During Early Development and in the Mature Animal," *Journal of Lipid Research* 45 (2004): 1375–1397.
183. S. Ando, Y. Tanaka, Y. Toyoda, and K. Kon, "Turnover of Myelin Lipids in Aging Brain," *Neurochemical Research* 28 (2003): 5–13.
184. D. S. Goodman and R. P. Noble, "Turnover of Plasma Cholesterol in Man," *Journal of Clinical Investigation* 47 (1968): 231–241.
185. H. Yoon, A. Kleven, A. Paulsen, et al., "Interplay Between Exercise and Dietary Fat Modulates Myelinogenesis in the Central Nervous System," *Biochimica et Biophysica Acta* 1862 (2016): 545–555.
186. J. Zheng, X. Sun, C. Ma, B. M. Li, and F. Luo, "Voluntary Wheel Running Promotes Myelination in the Motor Cortex Through Wnt Signaling in Mice," *Molecular Brain* 12 (2019): 85.
187. S. K. Jensen, N. J. Michaels, S. Ilyntskyy, M. B. Keough, O. Kovalchuk, and V. W. Yong, "Multimodal Enhancement of Remyelination by Exercise With a Pivotal Role for Oligodendroglial PGC1alpha," *Cell Reports* 24 (2018): 3167–3179.
188. L. Chen, F. L. Chao, W. Lu, et al., "Long-Term Running Exercise Delays Age-Related Changes in White Matter in Rats," *Frontiers in Aging Neuroscience* 12 (2020): 590530.
189. N. Feter, M. P. Freitas, N. G. Gonzales, D. Umpierre, R. K. Cardoso, and A. J. Rombaldi, "Effects of Physical Exercise on Myelin Sheath Regeneration: A Systematic Review and Meta-Analysis," *Science & Sports* 33 (2018): 8–21.
190. Q. Xiao, F. Wang, Y. Luo, et al., "Exercise Protects Myelinated Fibers of White Matter in a Rat Model of Depression," *Journal of Comparative Neurology* 526 (2018): 537–549.
191. F. L. Chao, L. Zhang, Y. Zhang, et al., "Running Exercise Protects Against Myelin Breakdown in the Absence of Neurogenesis in the Hippocampus of AD Mice," *Brain Research* 1684 (2018): 50–59.
192. A. Z. Burzynska, L. Chaddock-Heyman, M. W. Voss, et al., "Physical Activity and Cardiorespiratory Fitness Are Beneficial for White Matter in Low-Fit Older Adults," *PLoS One* 9 (2014): e107413.
193. B. Y. Tseng, T. Gundapuneedi, M. A. Khan, et al., "White Matter Integrity in Physically Fit Older Adults," *NeuroImage* 82 (2013): 510–516.
194. M. W. Voss, S. Heo, R. S. Prakash, et al., "The Influence of Aerobic Fitness on Cerebral White Matter Integrity and Cognitive Function in Older Adults: Results of a One-Year Exercise Intervention," *Human Brain Mapping* 34 (2013): 2972–2985.
195. T. Tarumi, B. P. Thomas, B. Y. Tseng, et al., "Cerebral White Matter Integrity in Amnesic Mild Cognitive Impairment: A 1-Year Randomized Controlled Trial of Aerobic Exercise Training," *Journal of Alzheimer's Disease* 73 (2020): 489–501.
196. J. M. Northey, N. Cherbuin, K. L. Pumpa, D. J. Smees, and B. Rattray, "Exercise Interventions for Cognitive Function in Adults Older Than 50: A Systematic Review With Meta-Analysis," *British Journal of Sports Medicine* 52, no. 3 (2017): 154–160, <https://doi.org/10.1136/bjsports-2016-096587>.
197. J. Ilha, R. T. Araujo, T. Malysz, et al., "Endurance and Resistance Exercise Training Programs Elicit Specific Effects on Sciatic Nerve Regeneration After Experimental Traumatic Lesion in Rats," *Neurorehabilitation and Neural Repair* 22 (2008): 355–366.
198. N. L. van Meeteren, J. H. Brakkee, P. J. Helden, and W. H. Gispen, "The Effect of Exercise Training on Functional Recovery After Sciatic Nerve Crush in the Rat," *Journal of the Peripheral Nervous System* 3 (1998): 277–282.
199. V. K. Venkatraman, A. Sanderson, K. L. Cox, et al., "Effect of a 24-Month Physical Activity Program on Brain Changes in Older Adults at Risk of Alzheimer's Disease: The AIBL Active Trial," *Neurobiology of Aging* 89 (2020): 132–141.
200. K. R. Norum, T. Berg, P. Helgerud, and C. A. Drevon, "Transport of Cholesterol," *Physiological Reviews* 63 (1983): 1343–1419.
201. A. S. Plump, J. D. Smith, T. Hayek, et al., "Severe Hypercholesterolemia and Atherosclerosis in Apolipoprotein E-Deficient Mice Created by Homologous Recombination in ES Cells," *Cell* 71 (1992): 343–353.
202. M. F. Linton, R. Gish, S. T. Hubl, et al., "Phenotypes of Apolipoprotein B and Apolipoprotein E After Liver Transplantation," *Journal of Clinical Investigation* 88 (1991): 270–281.
203. E. Masliah, M. Mallory, N. Ge, M. Alford, I. Veinbergs, and A. D. Roses, "Neurodegeneration in the Central Nervous System of apoE-Deficient Mice," *Experimental Neurology* 136 (1995): 107–122.
204. J. Poirier, "Apolipoprotein E in the Brain and Its Role in Alzheimer's Disease," *Journal of Psychiatry & Neuroscience* 21 (1996): 128–134.
205. M. Mato, S. Ookawara, T. Mashiko, et al., "Regional Difference of Lipid Distribution in Brain of Apolipoprotein E Deficient Mice," *Anatomical Record* 256 (1999): 165–176.
206. I. Soto, L. C. Graham, H. J. Richter, et al., "APOE Stabilization by Exercise Prevents Aging Neurovascular Dysfunction and Complement Induction," *PLoS Biology* 13 (2015): e1002279.
207. V. I. Zannis, J. L. Breslow, G. Utermann, et al., "Proposed Nomenclature of apoE Isoforms, apoE Genotypes, and Phenotypes," *Journal of Lipid Research* 23 (1982): 911–914.
208. R. W. Mahley and S. C. Rall, Jr., "Apolipoprotein E: Far More Than a Lipid Transport Protein," *Annual Review of Genomics and Human Genetics* 1 (2000): 507–537.
209. L. A. Farrer, "Effects of Age, Sex, and Ethnicity on the Association Between Apolipoprotein E Genotype and Alzheimer Disease," *JAMA* 278 (1997): 1349.
210. H. Zhang, P. W. Reymers, M. S. Liu, et al., "Patients With apoE3 Deficiency (E2/2, E3/2, and E4/2) who Manifest With Hyperlipidemia Have Increased Frequency of an Asn 291→Ser Mutation in the Human LPL Gene," *Arteriosclerosis, Thrombosis, and Vascular Biology* 15 (1995): 1695–1703.
211. P. Huebbe and G. Rimbach, "Evolution of Human Apolipoprotein E (APOE) Isoforms: Gene Structure, Protein Function and Interaction With Dietary Factors," *Ageing Research Reviews* 37 (2017): 146–161.

212. S. Egert, G. Rimbach, and P. Huebbe, "ApoE Genotype: From Geographic Distribution to Function and Responsiveness to Dietary Factors," *Proceedings of the Nutrition Society* 71 (2012): 410–424.
213. P. P. Singh, M. Singh, and S. S. Mastana, "APOE Distribution in World Populations With New Data From India and the UK," *Annals of Human Biology* 33 (2006): 279–308.
214. J. W. Blanchard, L. A. Akay, J. Davila-Velderrain, et al., "APOE4 Impairs Myelination via Cholesterol Dysregulation in Oligodendrocytes," *Nature* 611 (2022): 769–779.
215. G. Saher, S. Quintes, and K. A. Nave, "Cholesterol: A Novel Regulatory Role in Myelin Formation," *Neuroscientist* 17 (2011): 79–93.
216. D. C. Dean, 3rd, B. A. Jerskey, K. Chen, et al., "Brain Differences in Infants at Differential Genetic Risk for Late-Onset Alzheimer Disease: A Cross-Sectional Imaging Study," *JAMA Neurology* 71 (2014): 11–22.
217. Y. Yamazaki, N. Zhao, T. R. Caulfield, C. C. Liu, and G. Bu, "Apolipoprotein E and Alzheimer Disease: Pathobiology and Targeting Strategies," *Nature Reviews. Neurology* 15 (2019): 501–518.
218. J. M. Castellano, J. Kim, F. R. Stewart, et al., "Human apoE Isoforms Differentially Regulate Brain Amyloid-Beta Peptide Clearance," *Science Translational Medicine* 3 (2011): 89ra57.
219. M. K. Desai, K. L. Sudol, M. C. Janelins, M. A. Mastrangelo, M. E. Frazer, and W. J. Bowers, "Triple-Transgenic Alzheimer's Disease Mice Exhibit Region-Specific Abnormalities in Brain Myelination Patterns Prior to Appearance of Amyloid and Tau Pathology," *Glia* 57 (2009): 54–65.
220. L. Houdebine, C. A. Gallelli, M. Rastelli, N. K. Sampathkumar, and J. Grenier, "Effect of Physical Exercise on Brain and Lipid Metabolism in Mouse Models of Multiple Sclerosis," *Chemistry and Physics of Lipids* 207 (2017): 127–134.
221. M. Meissner, N. Nijstad, F. Kuipers, and U. J. Tietge, "Voluntary Exercise Increases Cholesterol Efflux but Not Macrophage Reverse Cholesterol Transport In Vivo in Mice," *Nutrition & Metabolism (London)* 7 (2010): 54.
222. C. Wei, M. Penumetcha, N. Santanam, Y. G. Liu, M. Garelnabi, and S. Parthasarathy, "Exercise Might Favor Reverse Cholesterol Transport and Lipoprotein Clearance: Potential Mechanism for Its Anti-Atherosclerotic Effects," *Biochimica et Biophysica Acta* 1723 (2005): 124–127.
223. K. Møllgaard and N. R. Saunders, "The Development of the Human Blood-Brain and Blood-CSF Barriers," *Neuropathology and Applied Neurobiology* 12 (1986): 337–358.
224. E. E. Goldmann, *Vitalfärbung am Zentralnervensystem: Beitrag zur Physio-Pathologie des Plexus chorioideus und der Hirnhäute* (Königl. Akademie der Wissenschaften, 1913).
225. E. M. Cornford, L. D. Braun, and W. H. Oldendorf, "Developmental Modulations of Blood-Brain Barrier Permeability as an Indicator of Changing Nutritional Requirements in the Brain," *Pediatric Research* 16 (1982): 324–328.
226. C. Parrado-Fernandez, K. Blennow, M. Hansson, V. Leoni, A. Cedazo-Minguez, and I. Björkhem, "Evidence for Sex Difference in the CSF/Plasma Albumin Ratio in ~20 000 Patients and 335 Healthy Volunteers," *Journal of Cellular and Molecular Medicine* 22 (2018): 5151–5154.
227. R. L. Chen, "Is It Appropriate to Use Albumin CSF/Plasma Ratio to Assess Blood Brain Barrier Permeability?," *Neurobiology of Aging* 32 (2011): 1338–1339.
228. I. C. M. Verheggen, J. J. A. de Jong, M. P. J. van Boxtel, et al., "Increase in Blood-Brain Barrier Leakage in Healthy, Older Adults," *Geroscience* 42 (2020): 1183–1193.
229. I. C. M. Verheggen, J. J. A. de Jong, M. P. J. van Boxtel, et al., "Imaging the Role of Blood-Brain Barrier Disruption in Normal Cognitive Ageing," *Geroscience* 42 (2020): 1751–1764.
230. M. G. Fragas, V. B. Candido, G. G. Davanzo, C. Rocha-Santos, A. Ceroni, and L. C. Michelini, "Transcytosis Within PVN Capillaries: A Mechanism Determining Both Hypertension-Induced Blood-Brain Barrier Dysfunction and Exercise-Induced Correction," *American Journal of Physiology. Regulatory, Integrative and Comparative Physiology* 321 (2021): R732–R741.
231. L. Buttler, M. T. Jordao, M. G. Fragas, A. Ruggeri, A. Ceroni, and L. C. Michelini, "Maintenance of Blood-Brain Barrier Integrity in Hypertension: A Novel Benefit of Exercise Training for Autonomic Control," *Frontiers in Physiology* 8 (2017): 1048.
232. H. A. Raquel, S. M. Perego, G. S. Masson, L. Jensen, A. Colquhoun, and L. C. Michelini, "Blood-Brain Barrier Lesion – A Novel Determinant of Autonomic Imbalance in Heart Failure and the Effects of Exercise Training," *Clinical Science (London, England)* 137 (2023): 1049–1066.
233. C. Brown, S. Pemberton, A. Babin, et al., "Insulin Blood-Brain Barrier Transport and Interactions Are Greater Following Exercise in Mice," *Journal of Applied Physiology (Bethesda, MD: 1985)* 132 (2022): 824–834.
234. A. L. Oblak, P. B. Lin, K. P. Kotredes, et al., "Comprehensive Evaluation of the 5XFAD Mouse Model for Preclinical Testing Applications: A MODEL-AD Study," *Frontiers in Aging Neuroscience* 13 (2021): 713726.
235. O. Razi, A. Parnow, I. Rashidi, N. Pakravan, S. E. Nedaei, and R. W. Motl, "Aerobic Training Improves Blood-Brain Barrier and Neuronal Apoptosis in Experimental Autoimmune Encephalomyelitis," *Iranian Journal of Basic Medical Sciences* 25 (2022): 245–253.
236. M. U. Chupel, L. G. Minuzzi, G. Furtado, et al., "Exercise and Taurine in Inflammation, Cognition, and Peripheral Markers of Blood-Brain Barrier Integrity in Older Women," *Applied Physiology, Nutrition, and Metabolism* 43 (2018): 733–741.
237. Z. Zhu, J. Xu, Y. Jin, L. Wang, and X. Li, "Effects of Aerobic Exercise on Markers of Brain Injury in Methamphetamine-Dependent Individuals: A Randomized Controlled Trial," *Brain Sciences* 12, no. 11 (2022): 1521.
238. M. Savikj and J. R. Zierath, "Train Like an Athlete: Applying Exercise Interventions to Manage Type 2 Diabetes," *Diabetologia* 63 (2020): 1491–1499.
239. A. Toval, P. Solis-Urra, E. A. Bakker, et al., "Exercise and BRAIN Health in Patients With Coronary Artery Disease: Study Protocol for the HEART-BRAIN Randomized Controlled Trial," *Frontiers in Aging Neuroscience* 16 (2024): 1437567.